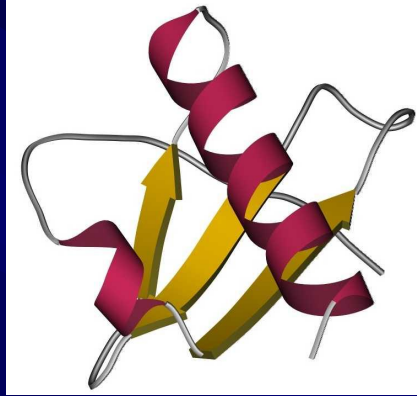


PROTEIN PHYSICS

LECTURES 11-12

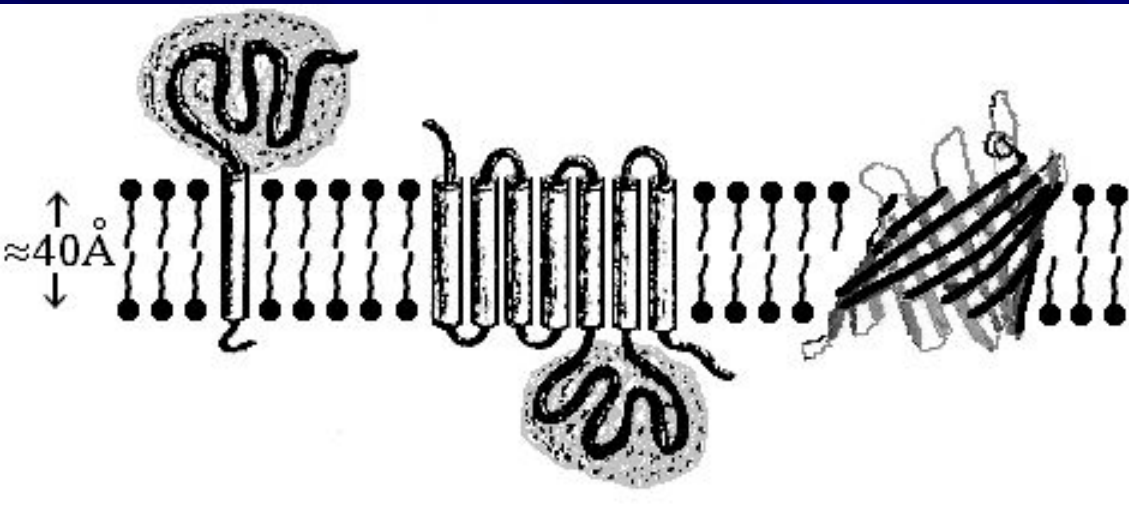
- Fibrous proteins and their functions
 - Membrane proteins and their functions
-
- Fibrous proteins: building blocks
 - Membrane proteins: transmitters



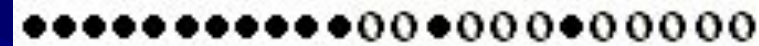
Globular proteins



quasi-random



Membrane proteins

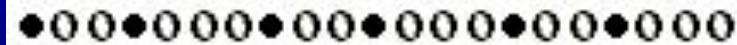


Hydro-
phobic
block

Hydro-
philic
block



Fibrous proteins

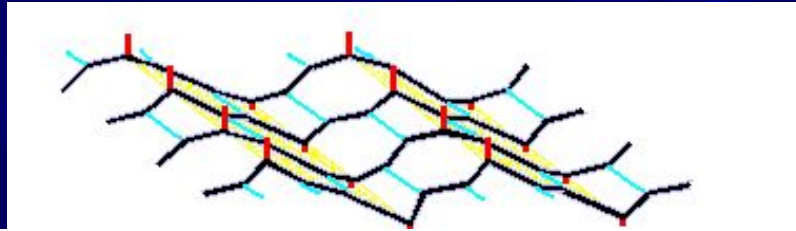


repeat | repeat |

H-bonds (NH:::OC) & hydrophobic forces

Fibrous proteins: regular building blocks

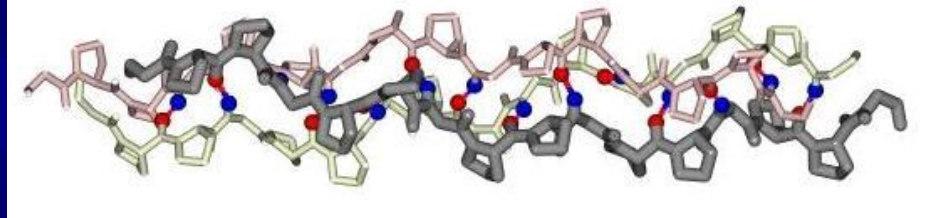
β



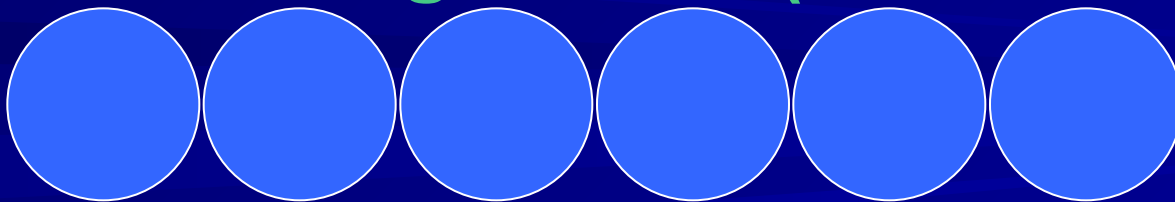
α

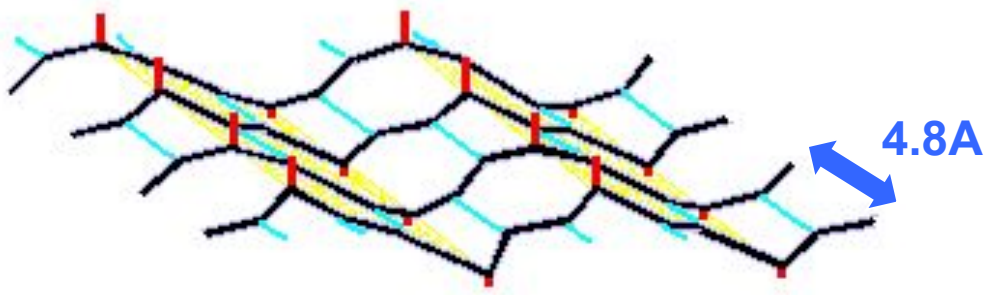


collagen



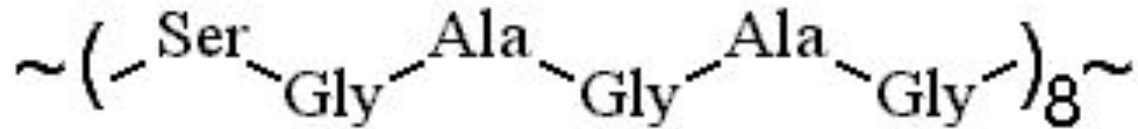
Here, we will not consider fibrous proteins
made of globules (actin, etc.)



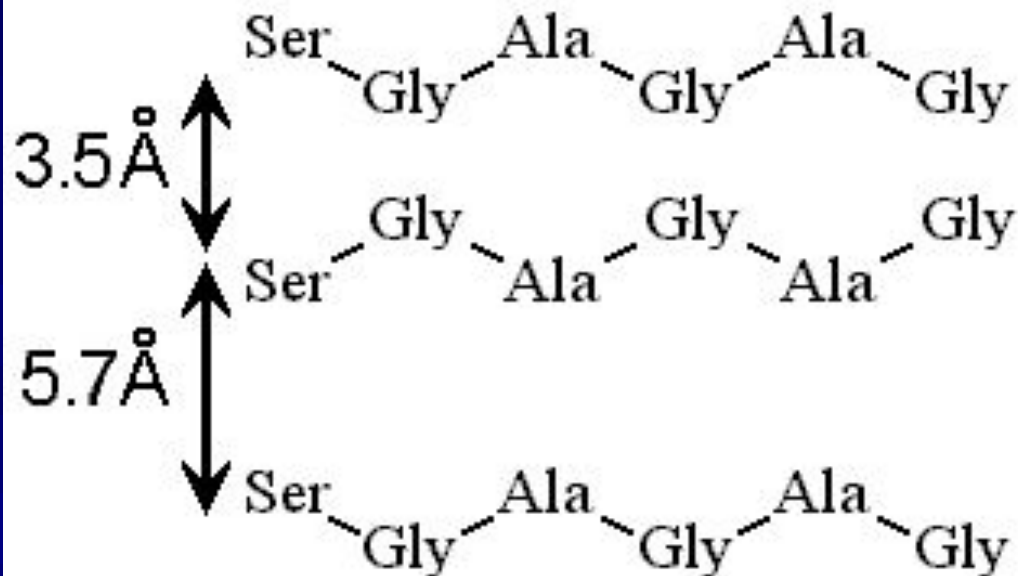


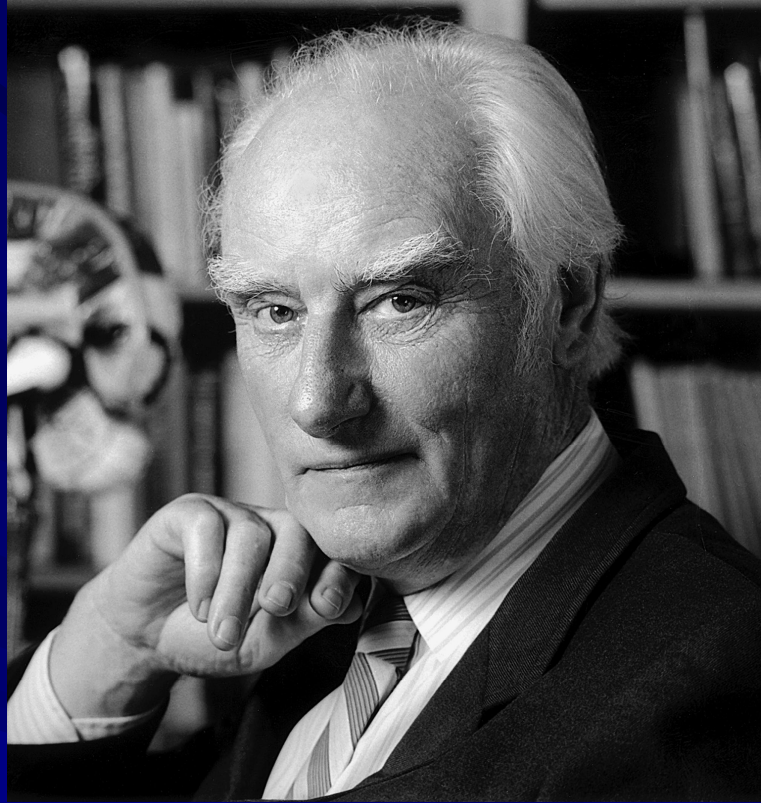
β

Silk fibroin



$\times \sim 50$



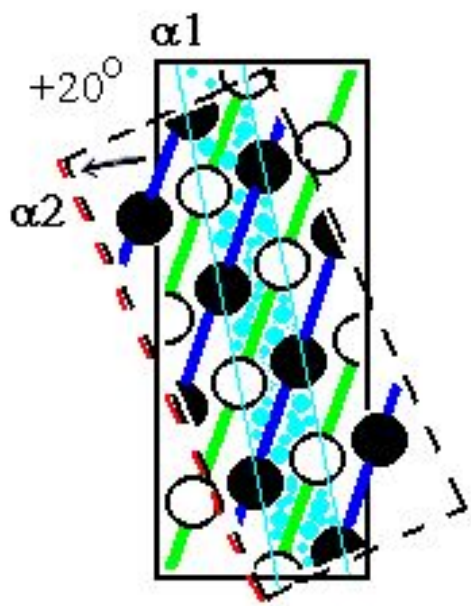
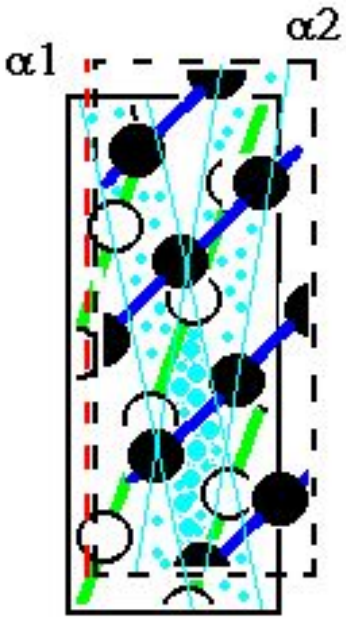
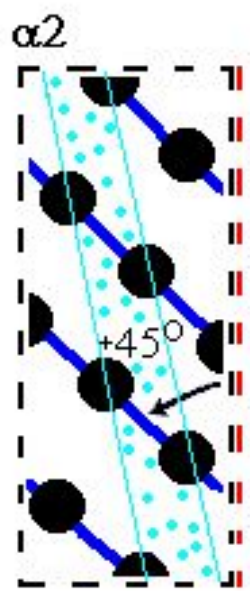
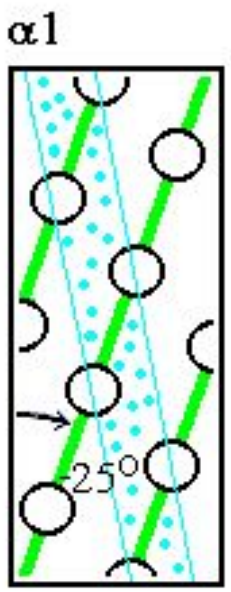
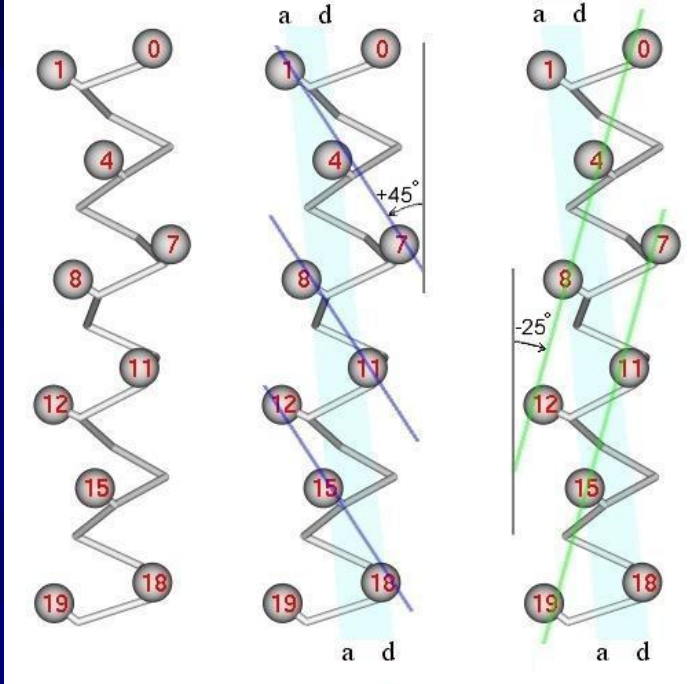
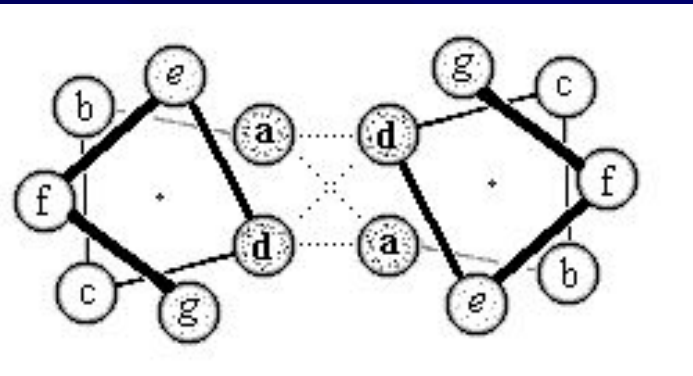


Francis Harry Compton **Crick** (1916 – 2004)
Nobel Prize 1962
for **DNA structure, 1953**

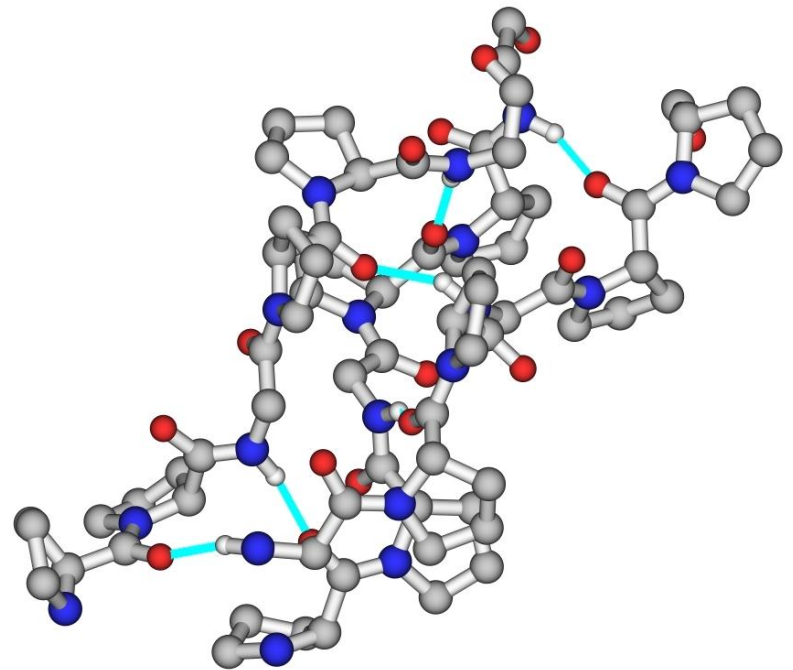
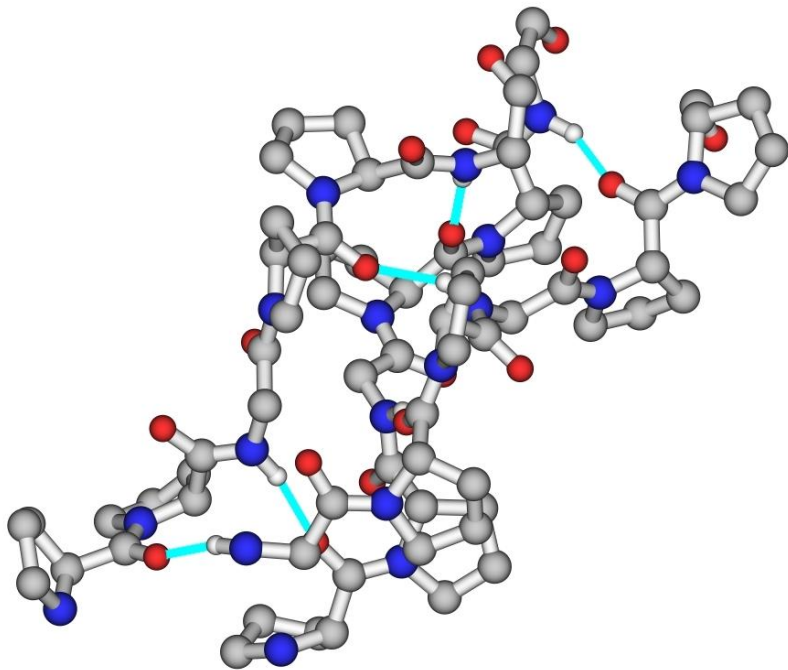
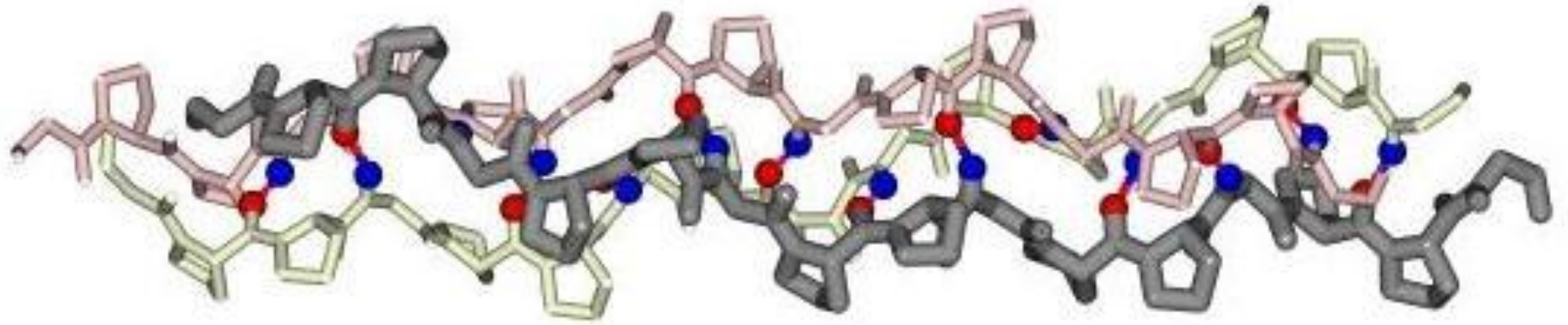
Coiled coil structure: F. Crick, 1952

C. Chothia, M. Levitt, D. Richardson, 1977

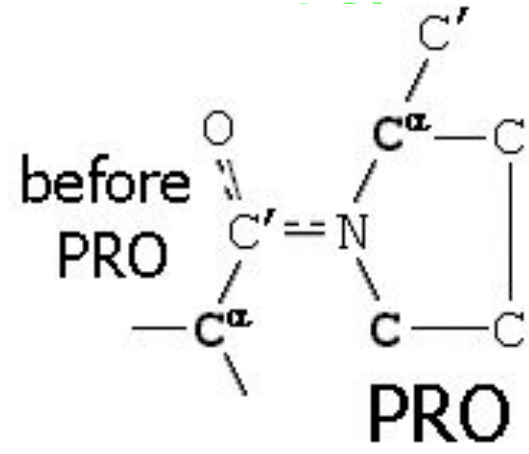
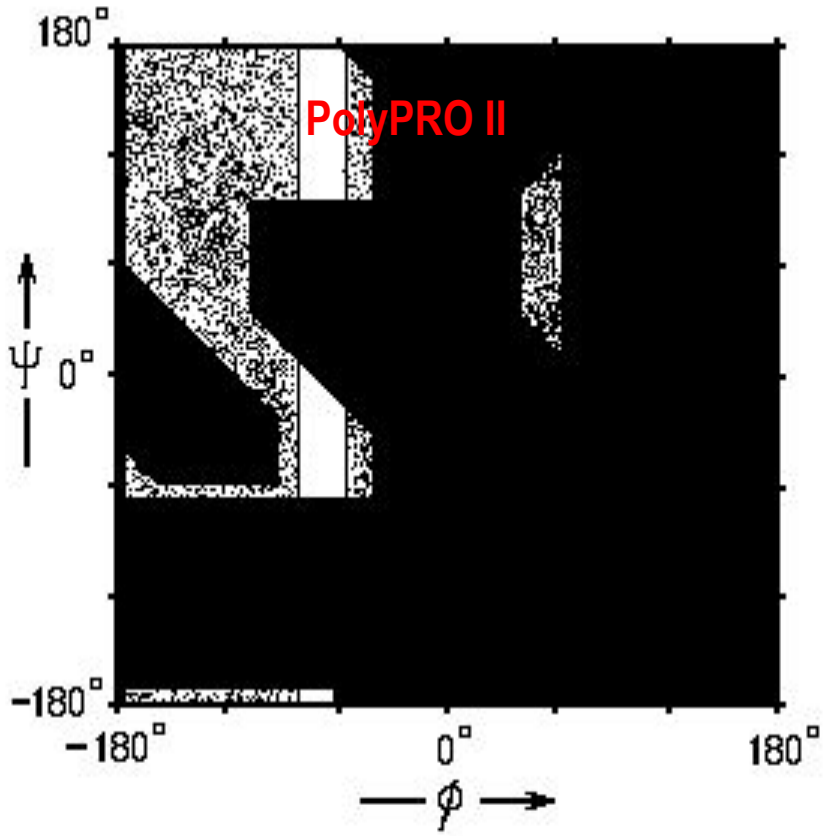
α -helix packing



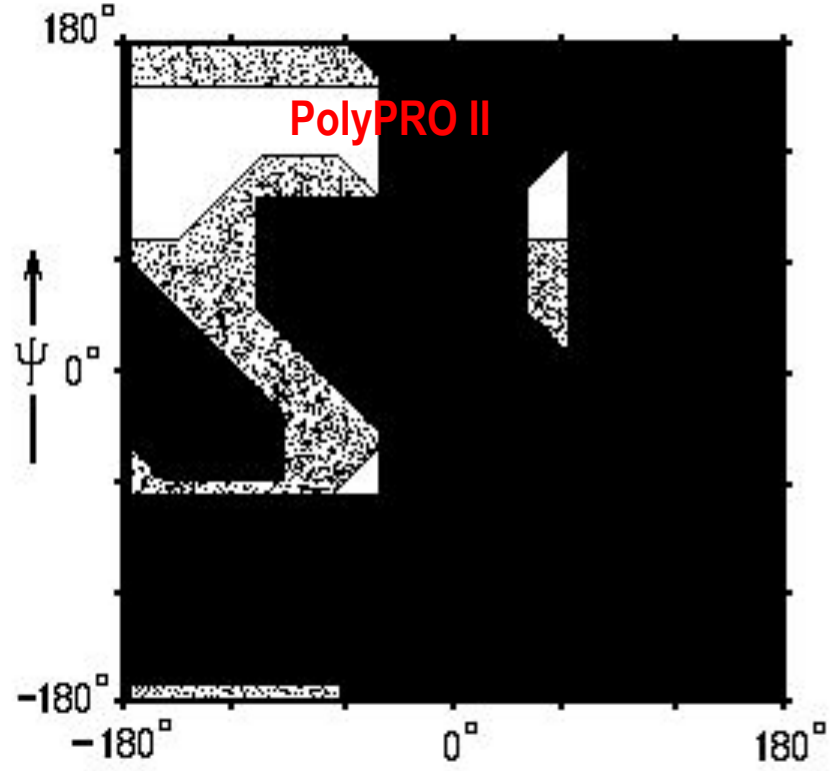
collagen triple helix: 3 chains \approx [Gly-X-Pro]_{~500}



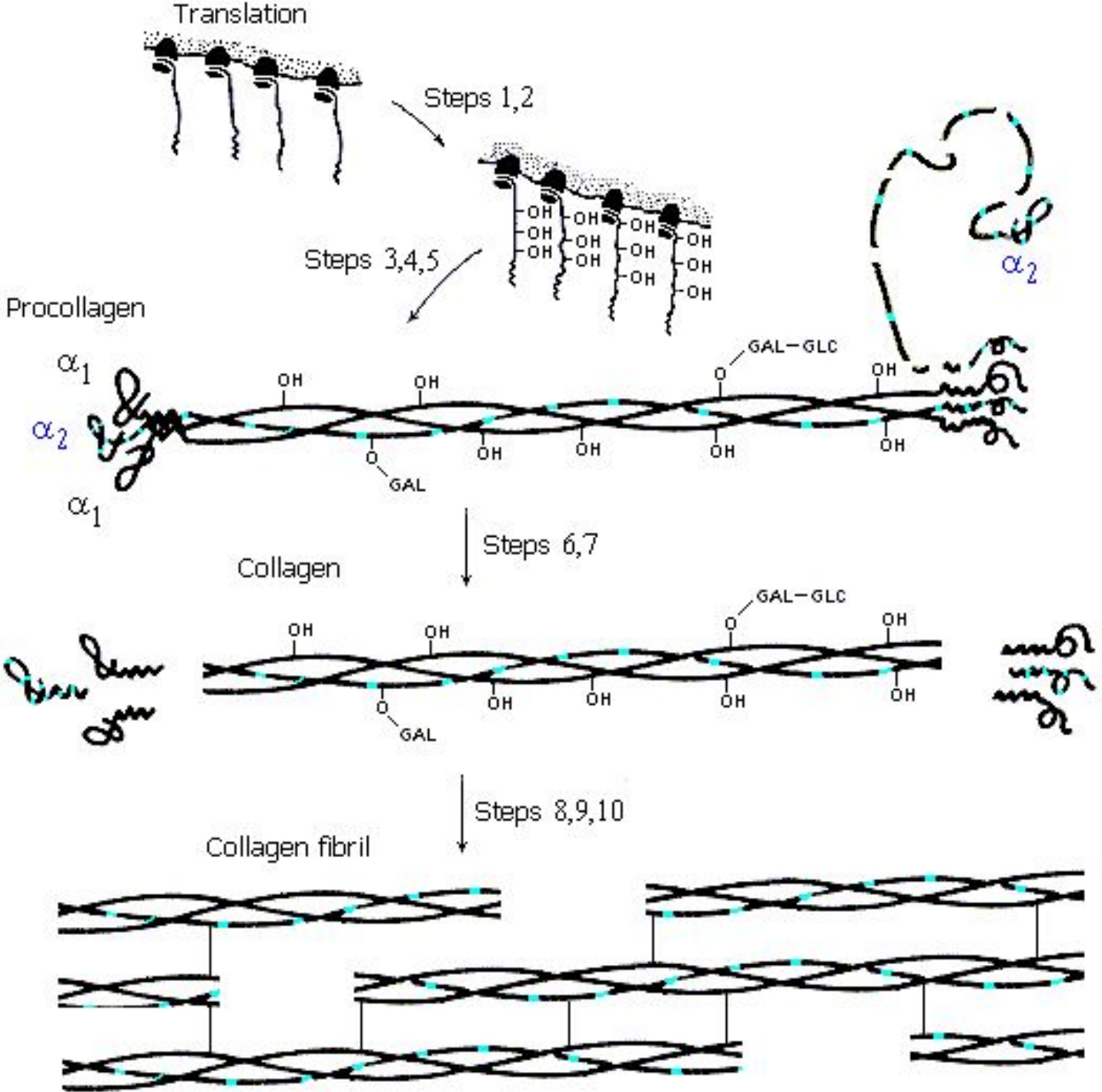
PRO ($\phi =$



Before PRO



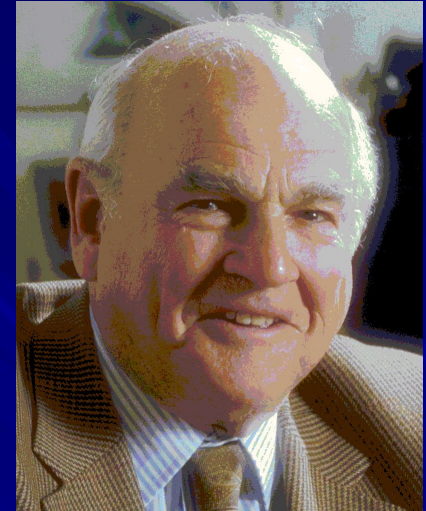
Collagen: assisted folding





Kuru: a mysterious disease, later demonstrated to be infectious prion disease.

Daniel Carleton **Gajdusek** (1923 –2008)
Baruch Samuel **Blumberg** (1925 – 2011)
Nobel Prize 1976



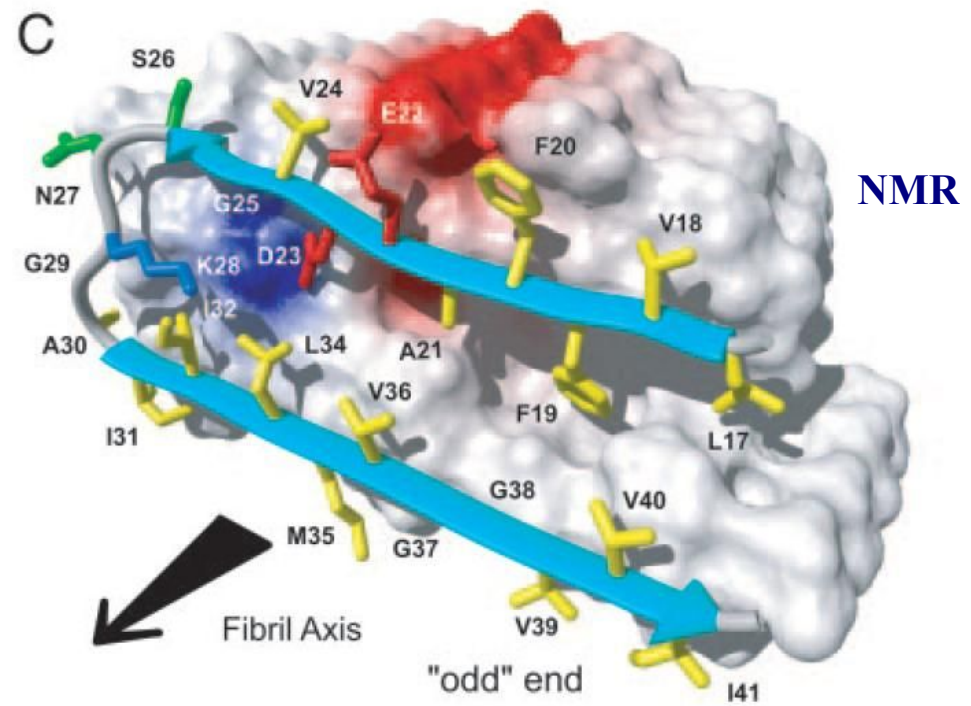
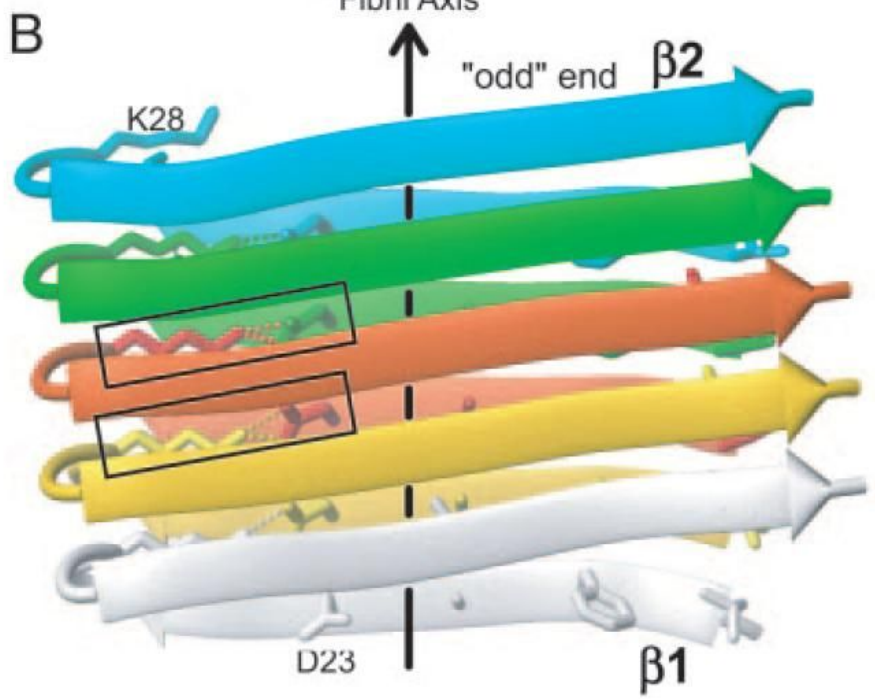
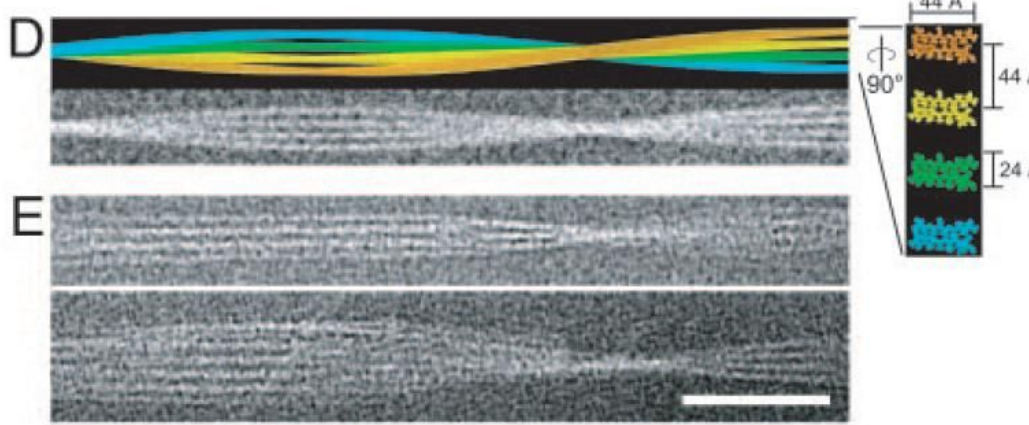
PRION: PRO*tein* and **In***fection*

Stanley Benjamin **Prusiner**, 1942
Nobel Prize 1997



Studies of amyloid formation

Christopher Martin **Dobson**, 1949
Royal Medal 2009

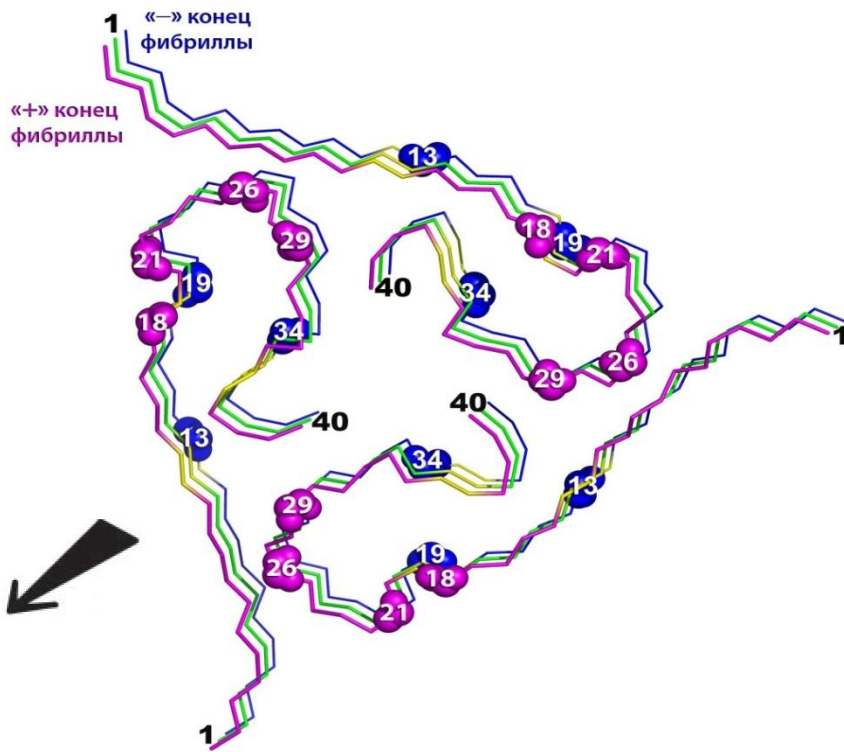


3D structure of Alzheimer's amyloid- β (1-42) fibrils

T.Lührs, C.Ritter, M.Adrian, D.Riek-Loher, B.Bohrmann, H.Döbeli, D.Schubert, R.Riek. *PNAS* 102:17342-17347 (2005)

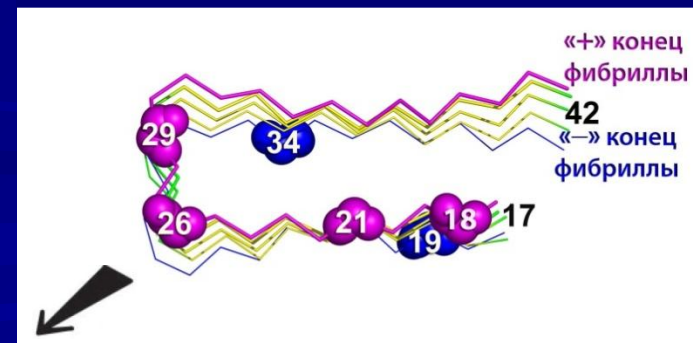


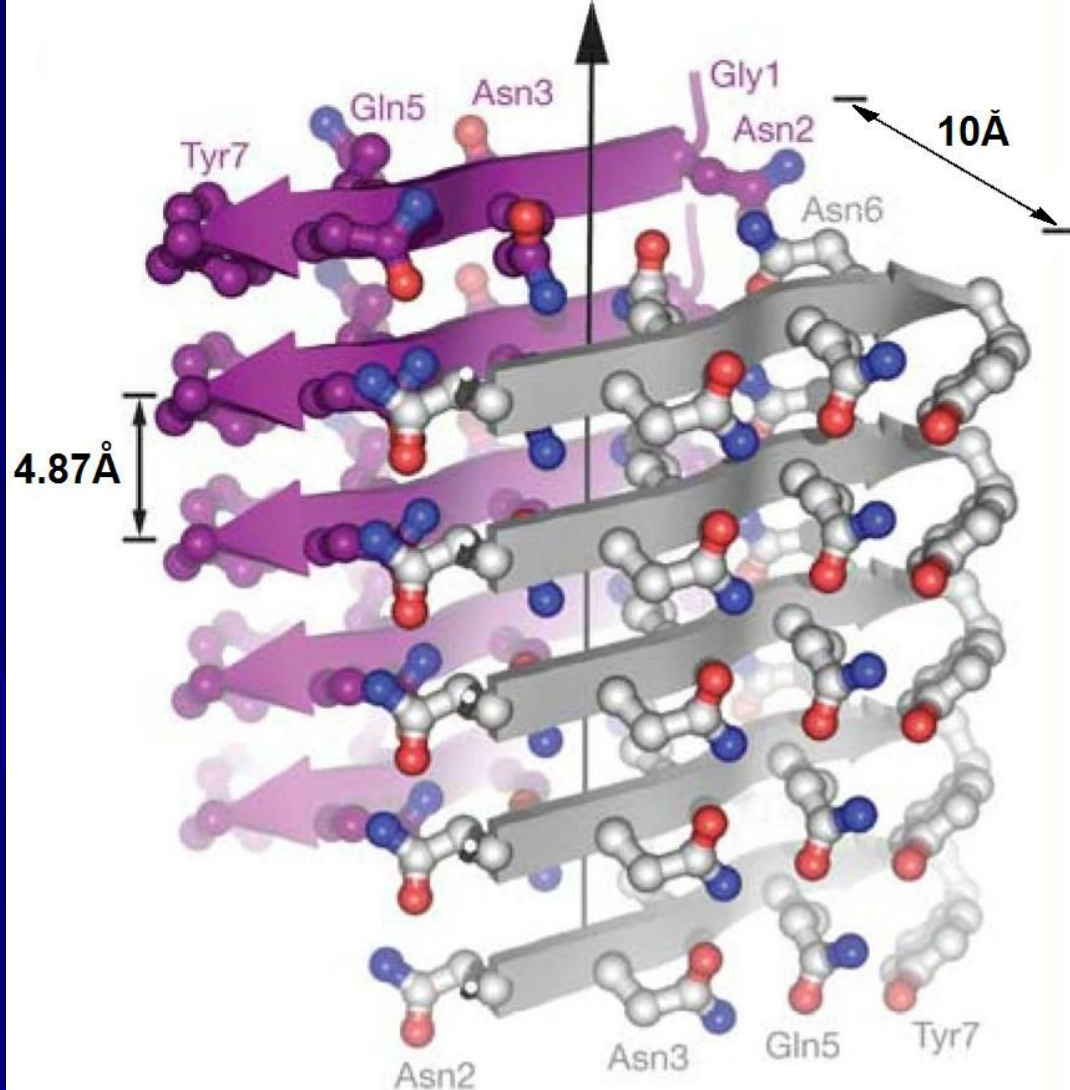
VARIABILITY OF STRUCTURES



Lührs T., Ritter C., Adrian M., Riek-Loher D., Bohrmann B., Döbeli H., Schubert D., Riek R.
3D structure of Alzheimer's amyloid-beta(1-42) fibrils.
PNAS 102:17342-17347 (2005).

Lu J.X., Qiang W., Yau W.M., Schwieters C.D., Meredith S.C., Tycko R.
Molecular structure of β -amyloid fibrils in Alzheimer's disease **brain tissue**.
Cell 154:1257-1268 (2013).

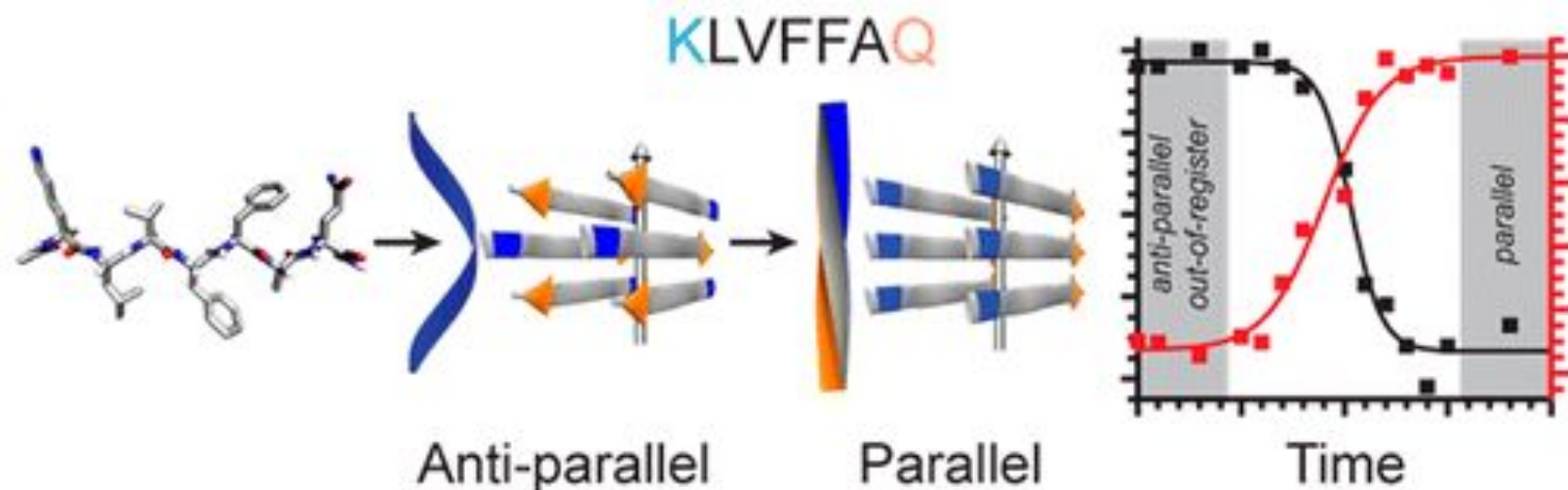




X-RAY

Structure of the cross- β spine of amyloid-like fibrils

R.Nelson, M.R.Sawaya, M.Balbirnie,
A.Ø.Madsen, C.Riek, R.Grothe, D.Eisenberg
Nature **435**:773-778 (2005)



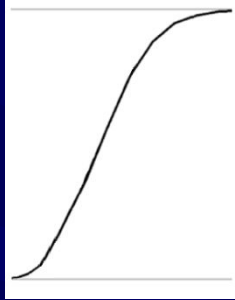
In contrast to an expected Ostwald-like ripening of amyloid assemblies, the nucleating core of the Dutch mutant of the A β peptide of Alzheimer's disease assembles through a series of conformational transitions. Structural characterization of the intermediate assemblies by isotope-edited IR and solid-state NMR reveals unexpected strand orientation intermediates and suggests new nucleation mechanisms in a progressive assembly pathway.

JACS → [Volume 136, Issue 43](#) → **October 29, 2014**¶

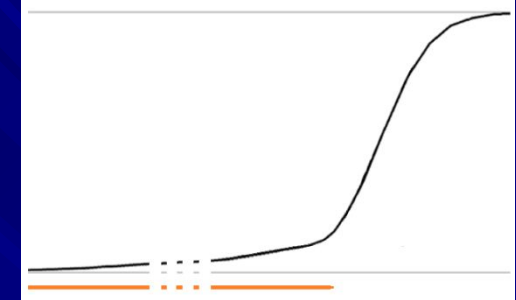
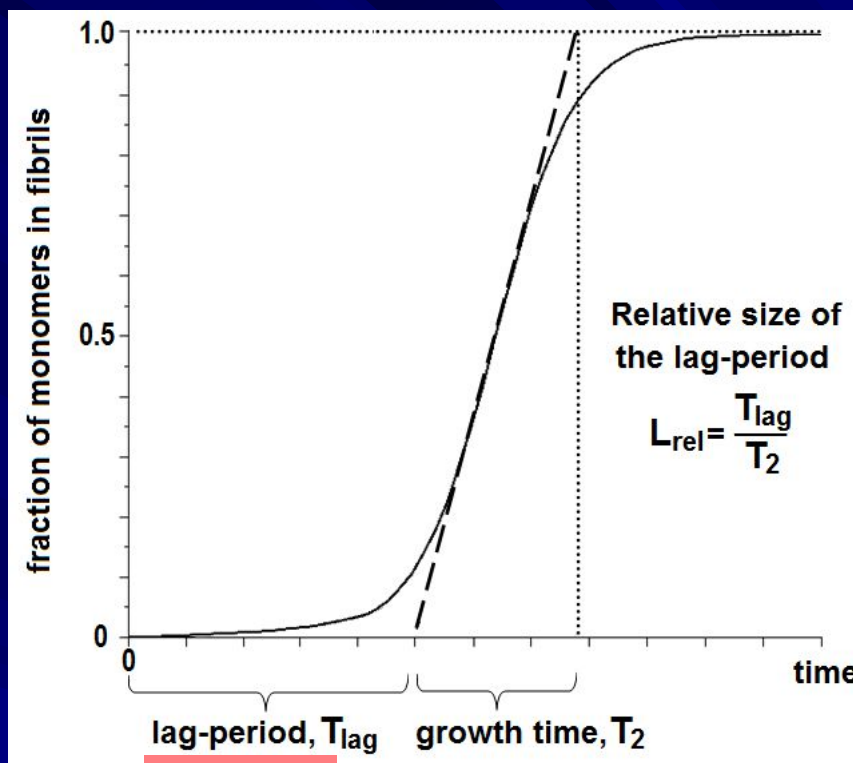
Kinetic Intermediates in Amyloid Assembly¶

Chen Liang, Rong Ni, Jillian E. Smith, W. Seth Childers, Anil K. Mehta, and David G. Lynn

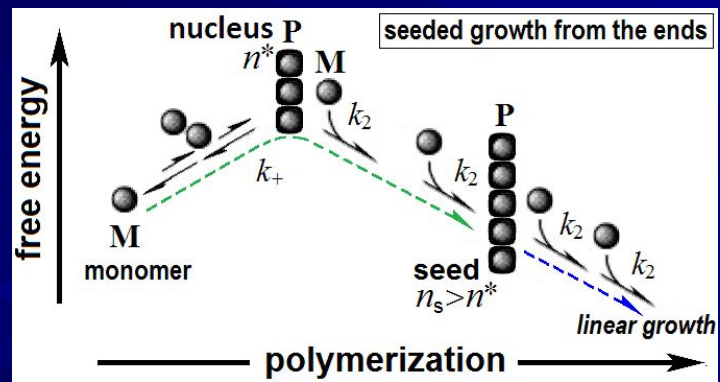
Growth of amyloids



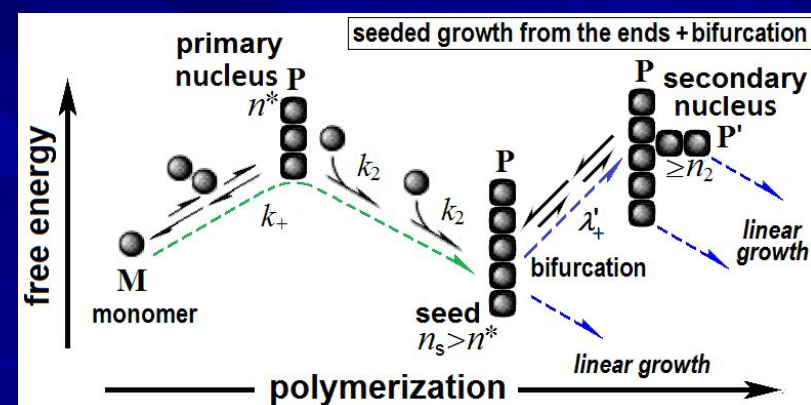
LINEAR GROWTH
NO LAG



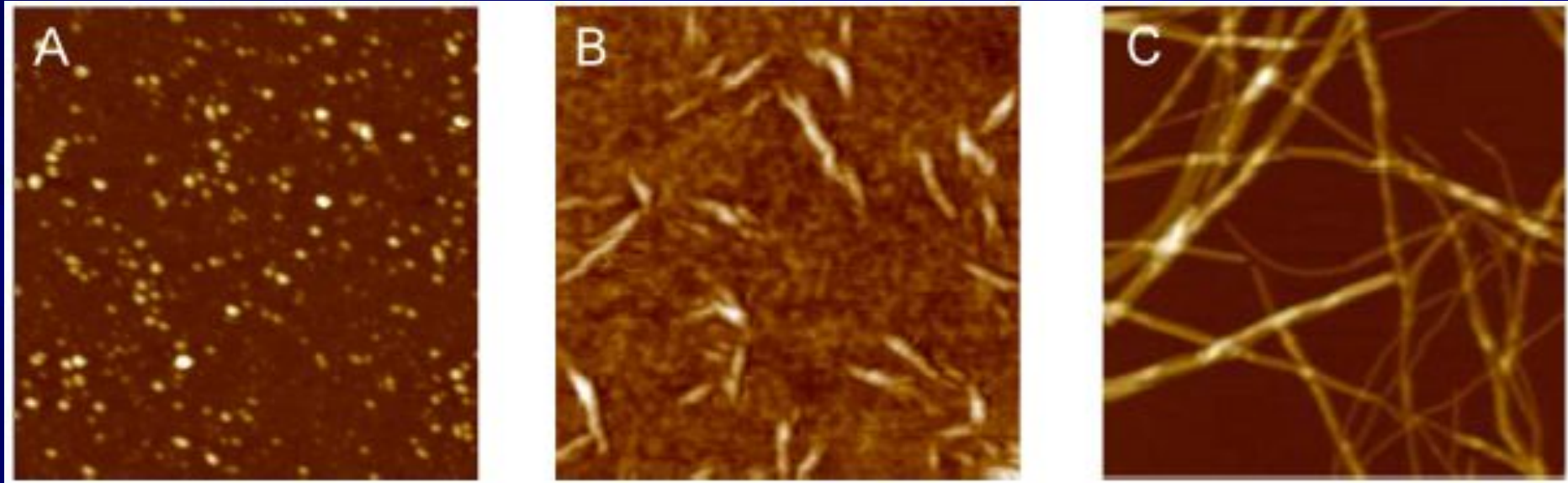
EXPONENTIAL GROWTH
VERY LARGE LAG



Different relative lag-period



Dovidchenko N.V., Finkelstein A.V., Galzitskaya O.V. 2014.
How to determine the size of folding nuclei of protofibrils from the concentration dependence of the rate and lag-time of aggregation. I. Modeling the amyloid protofibril formation.
J. Phys. Chem. B., 118:1189-1197.



Oligomers

Protofibrils

Mature amyloid fibrils

Atomic force microscopy

Relini A., Marano N., Gliozzi A. 2014.

Misfolding of amyloidogenic proteins and their interactions with membranes

Biomolecules, 4, 20-55 .

Natively non-structured fibrous proteins:

Elastin:

Matrix protein.

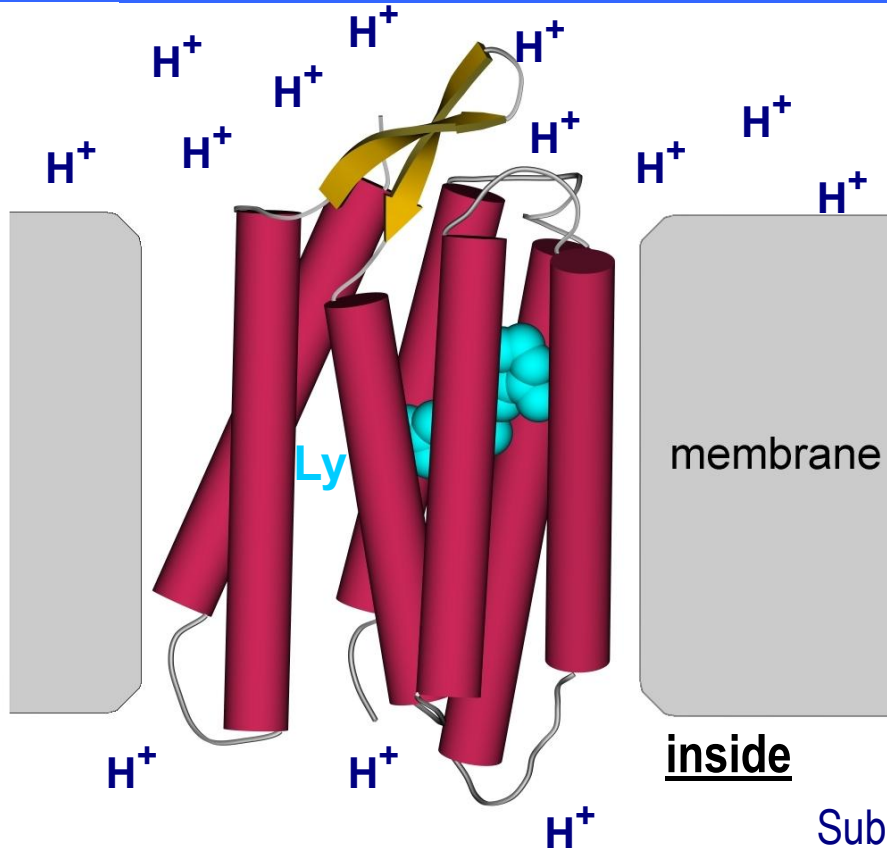
Short repeats.

Poor secondary structure.

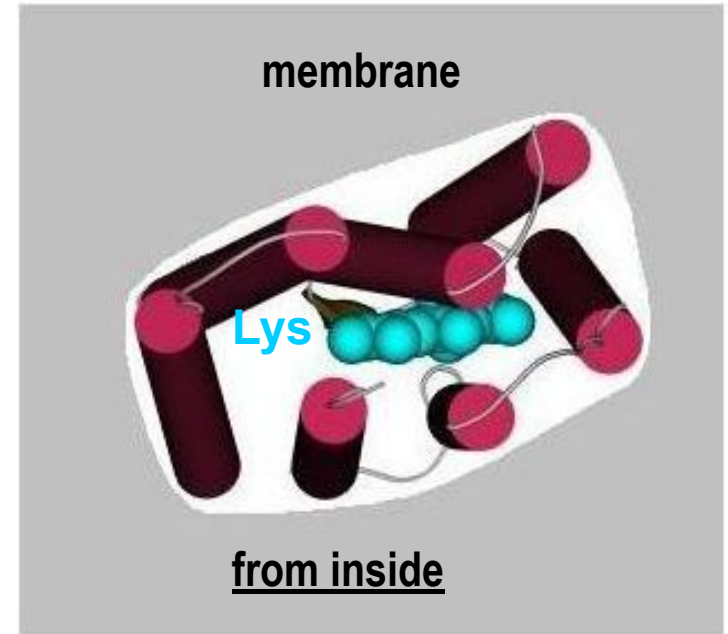
Chains are linked by chemically
modified Lys residues.

Like in rubber.

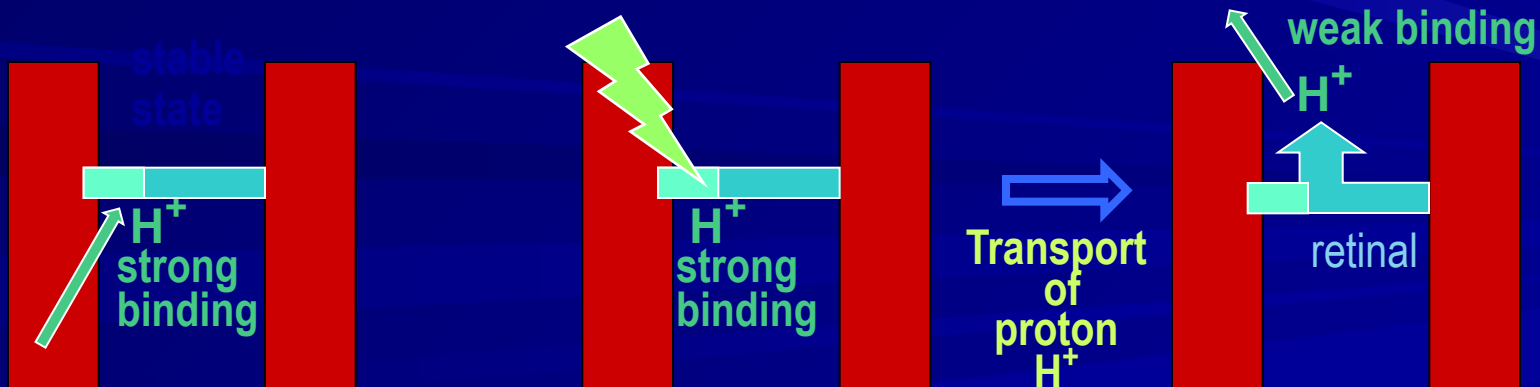
Bacteriorhodopsin (α) with retinal: the simplest transporter machine with a light-induced conformational change



Bacteriorhodopsin-Lys-retinal

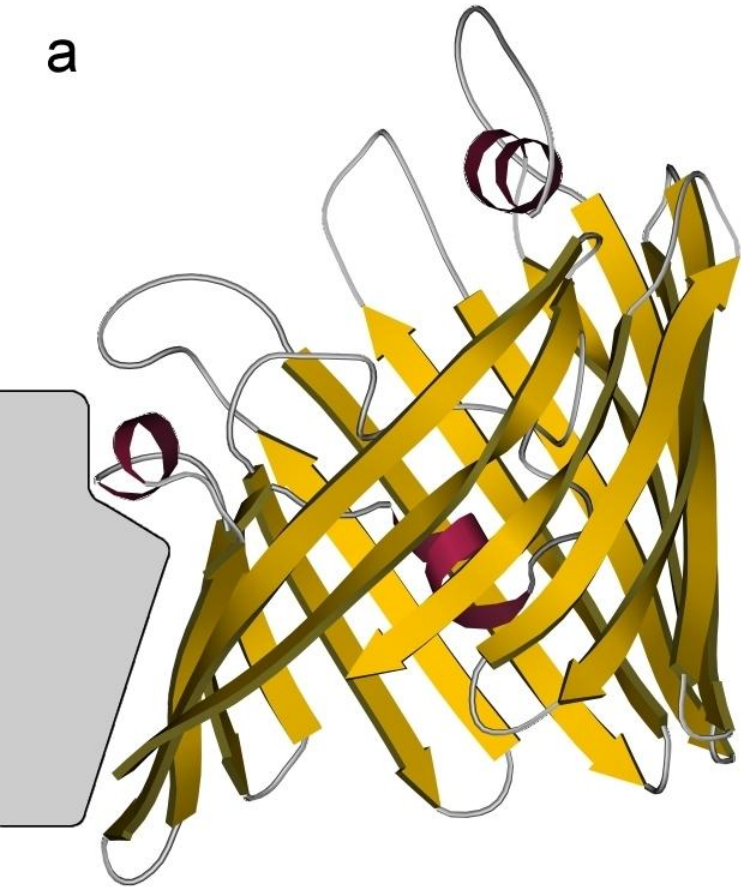


Subramaniam & Henderson, Nature 406, 653 (2000)



β

a



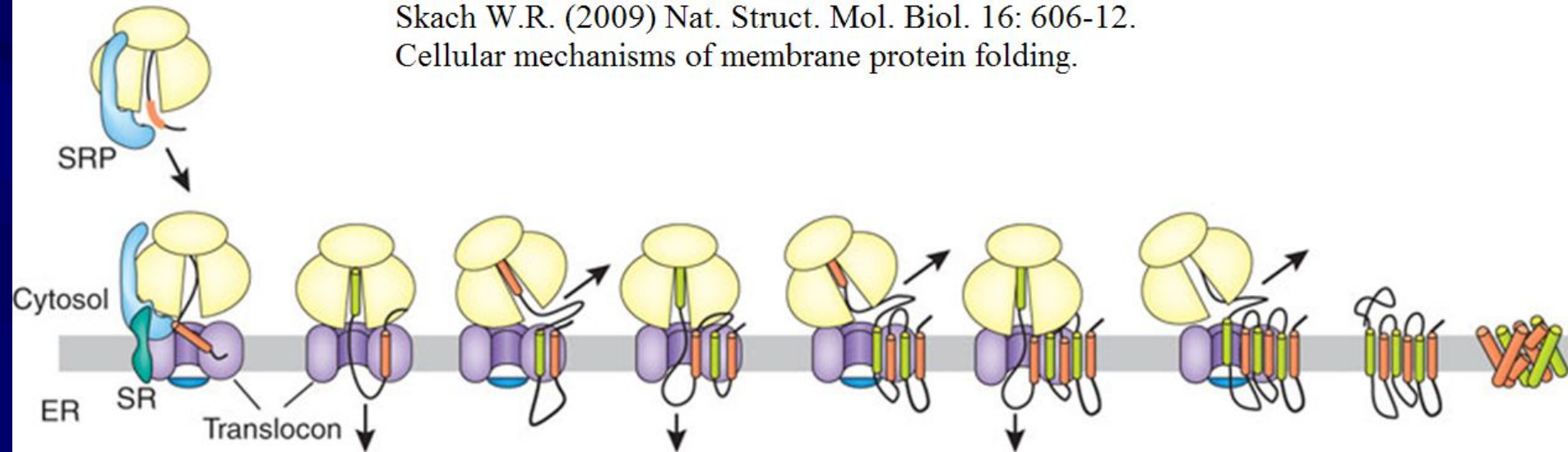
b



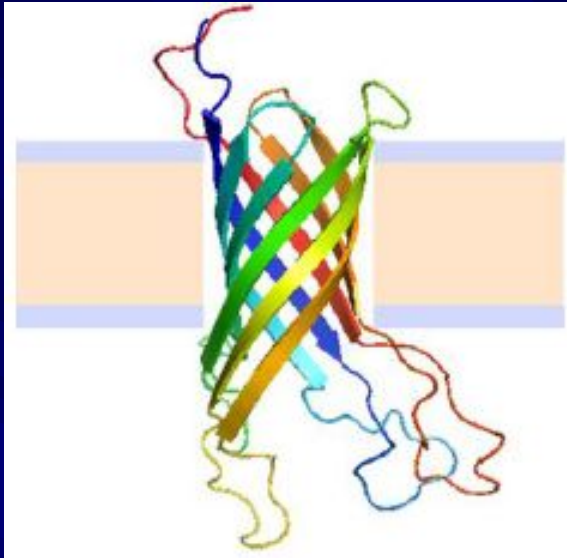
Porin
Transport of polar molecules

Membrane protein *in vivo*: Folding is assisted by “directing factors” - chaperones

Skach W.R. (2009) Nat. Struct. Mol. Biol. 16: 606-12.
Cellular mechanisms of membrane protein folding.



MANY OF **SIMPLE** MEMBRANE PROTEINS REFOLD *IN VITRO*
IN THE PRESENCE OF PHOSPHOLIPID VESICLES OR SURFACTANT MICELLES



COLLAPSED STATE: MIX OF COIL, α , β



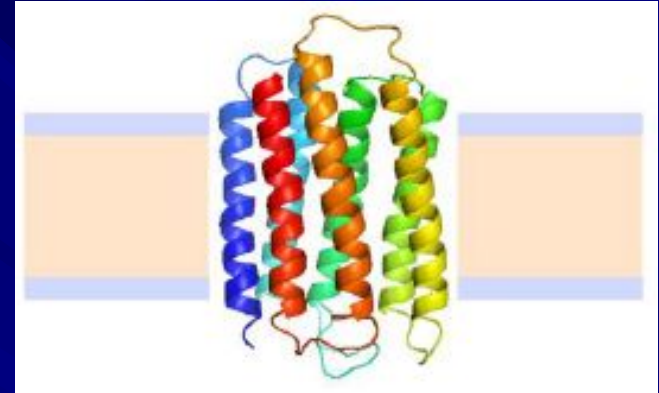
ASSOCIATES WITH **LIPID VESICLES**, β



DEEPER PENETRATION INTO LIPIDS



FULLY FOLDED



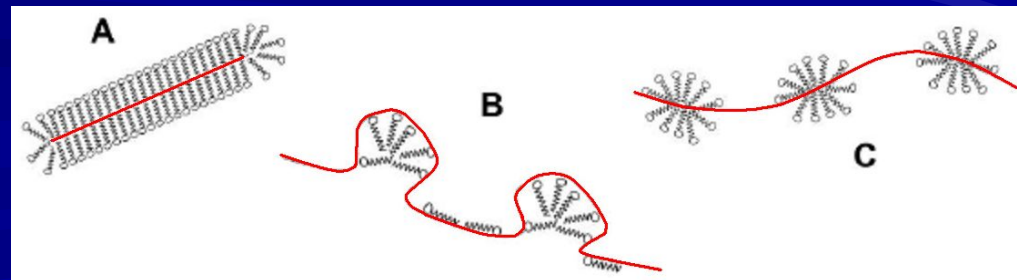
INDEPENDENT α -HELICES

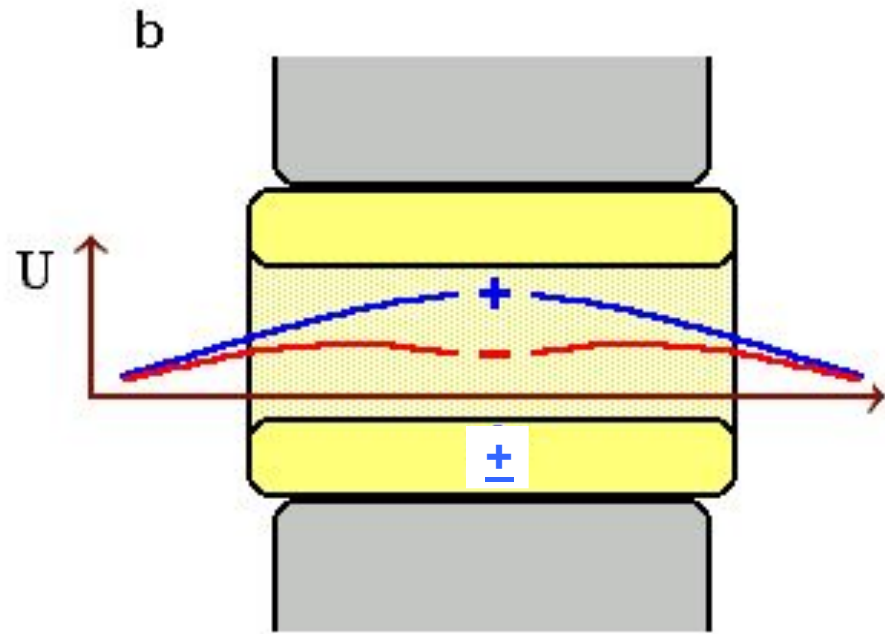
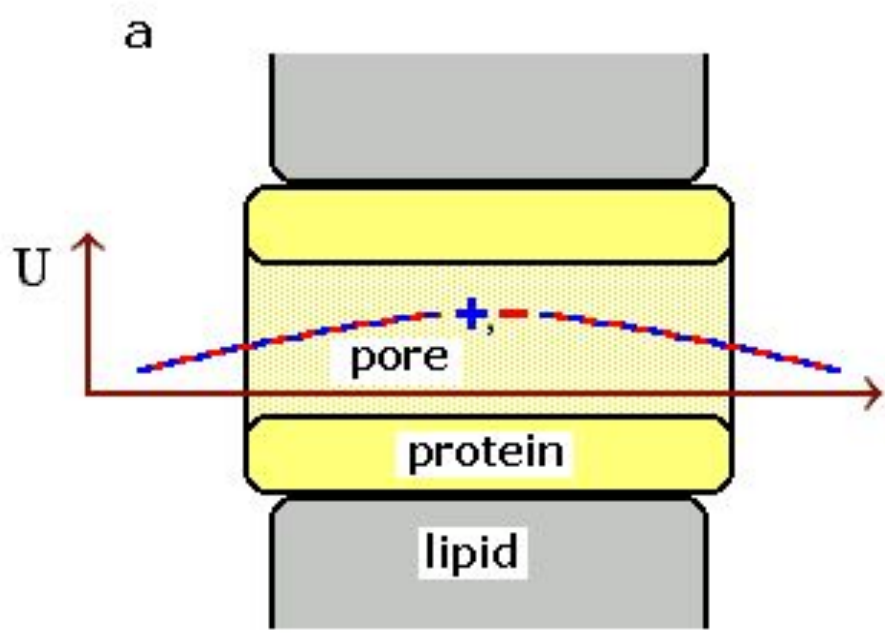


ASSEMBLE IN LIPID TO FULLY FOLDED

DIFFICULT TO STUDY:

DENATURED STATES OF MEMBRANE
PROTEINS ARE **DIVERSE & COMPLICATED**





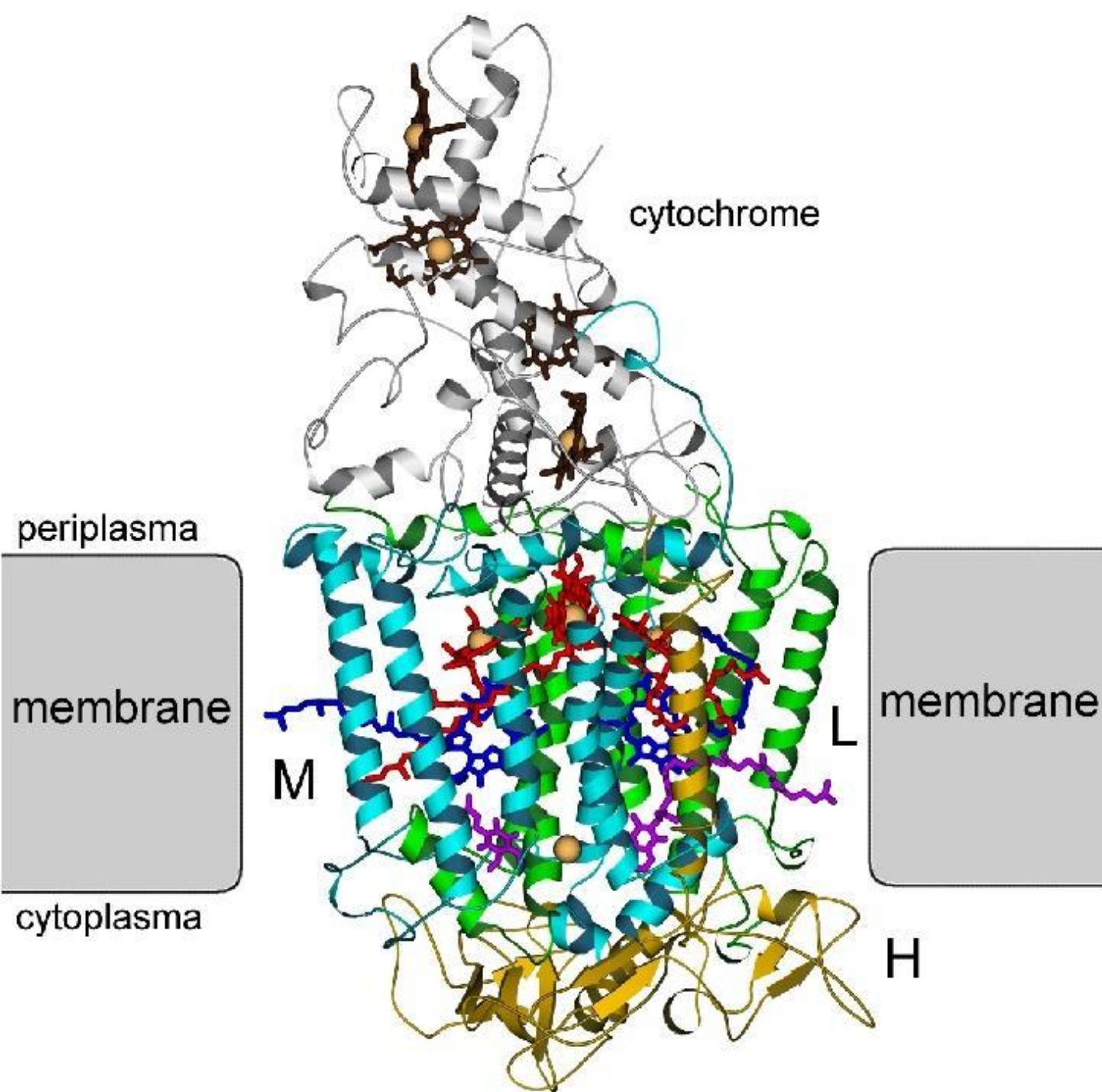
Pore in membrane: SELECTIVITY

Free energy of a charge in the non-charged non-polar pore:

$$\sim q^2 / [(\epsilon_{\text{MEMBR}} \epsilon_{\text{WATER}})^{1/2} r_{\text{PORE}}] \sim$$

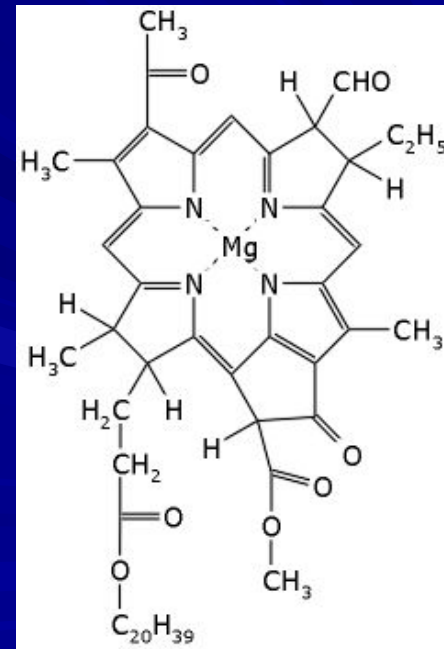
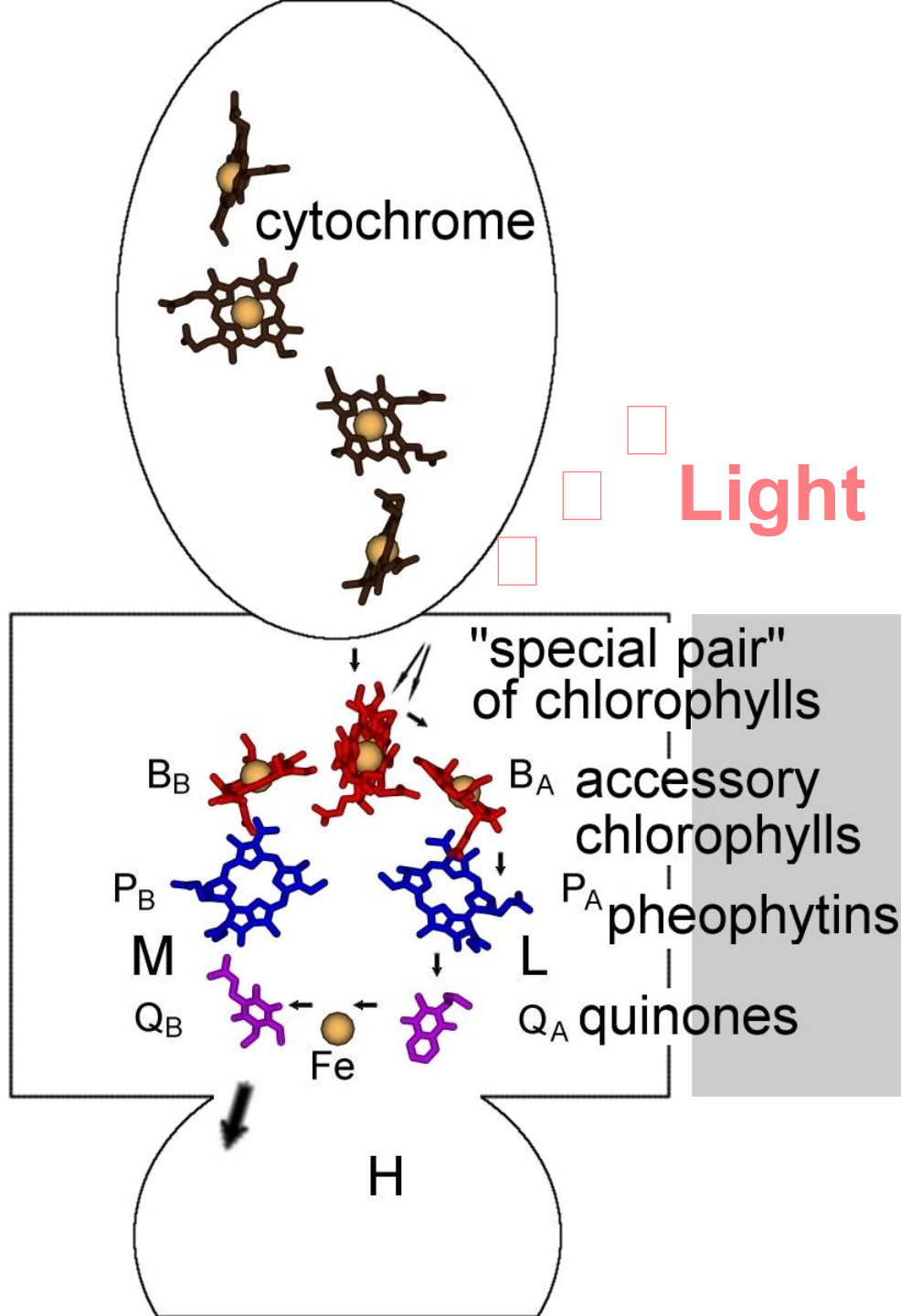
$$\sim 20 \text{ kcal/mol} / r_{\text{PORE}} (\text{\AA})$$

Photo-synthetic center



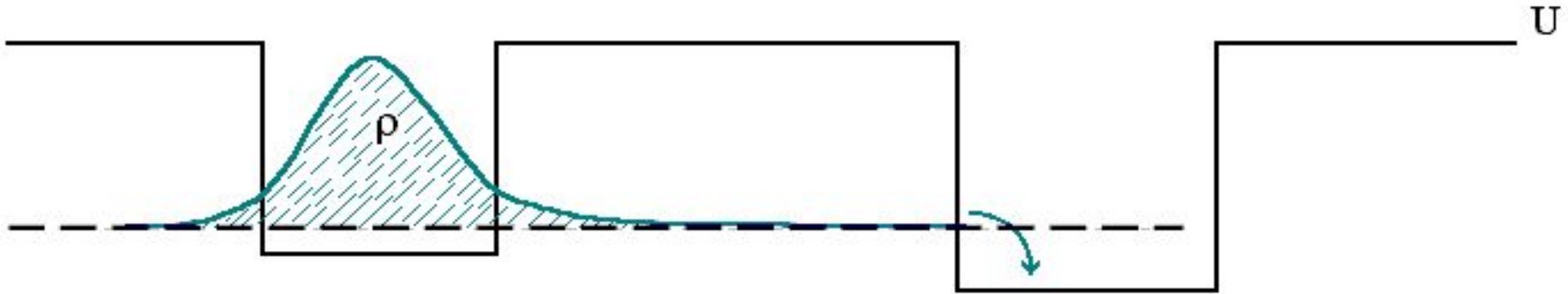
Robert Huber,
1937.
Nobel prize 1988

Pigments in photosynthetic center: Electron transfer



chlorophyll

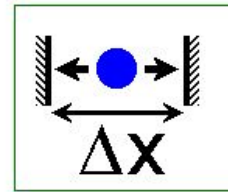
Tunneling



Atom $\approx 1\text{\AA}$ \Rightarrow Attenuation of electron density: $P(X) \sim 10^{-X(\text{\AA})}$

Heisenberg's uncertainty:

$$\Delta(mv) \cdot \Delta x \cong \hbar \text{ Planck's const}$$



$$v = \pm |v|$$

Energy of localization in Δx :

$$E = mv^2/2 \sim (\hbar^2/m)/(\Delta x^2)$$

DELOCALIZATION LEADS TO MORE STABLE STATE OF e

T-independent

Frequency of vibrations (attacks):

$$f \sim 10^{15}/\text{sec}$$

Successful attacks:

$$f_{\text{SUCCS.}}(x) \sim P(x) \cdot f, \text{ e.g.:}$$

$$f_{\text{SUCCS.}}(5\text{\AA}) \sim 10^{-5+15} \sim 10^{10}/\text{sec}$$