

Centrifugal Ultrafiltration Devices for.....

**protein / nucleic acid
concentration,
desalting, buffer
exchange** *and
more.....*



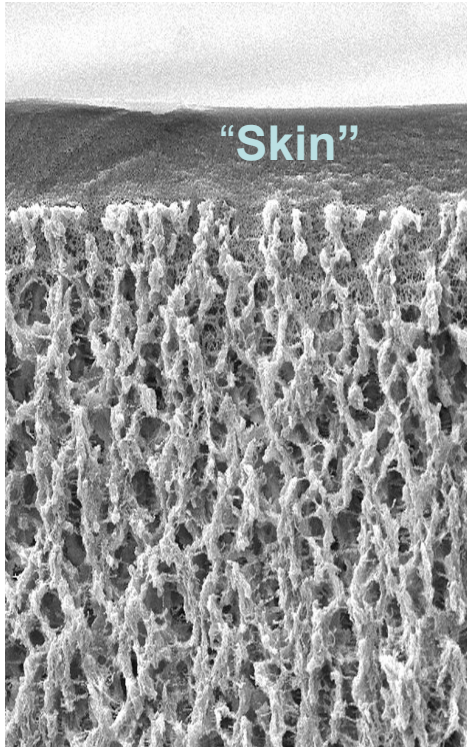
What is Ultrafiltration?

A process for the **separation** and/or **concentration of dissolved macromolecules** using an ultra-filtration membrane with a defined cut off

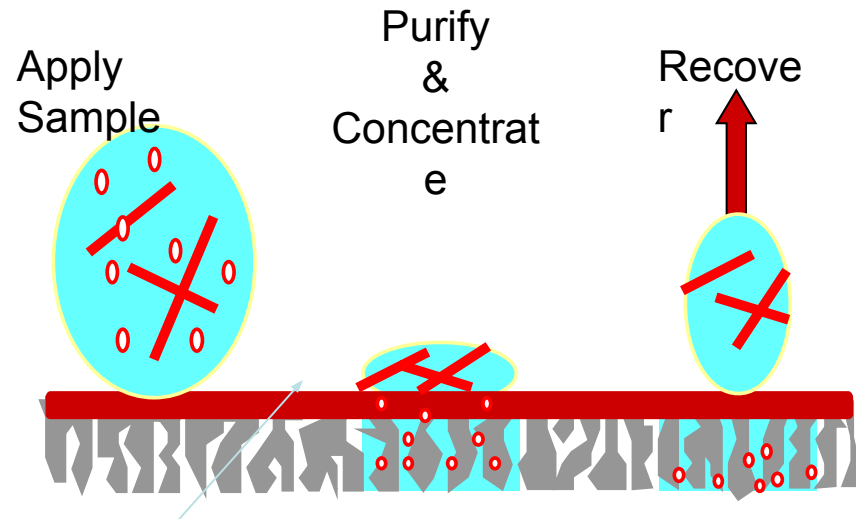
- Primary basis for separation is **molecular weight/size**
- **Driving force** can be **Centrifugal, Vacuum** or **Pressure**

What is an Ultrafiltration Membrane?

Structure of Ultrafiltration Membrane



Porous support



The UF 'skin' drives the separation

Ultracel® YM & PL regenerated cellulose

- Dense top layer = ultrafiltration membrane,
- Highly retentive
- Lowest protein binding
- Recommended for proteins; ideal for dilute proteins

How do I select the most suitable device?

- The **molecular weight** and or composition of the sample **determines NMWL (MWCO) cut off of the membrane**
- The **volume** and required concentration level of the sample determines **device** selection.
- The **selected device** determines the type of **rotor** and **G force**.

How do I select the most suitable membrane (cut-off) for my protein?

“Molecular Weight of Protein” divided by 2 or 3 = best cut-off of the ultrafiltration membrane

Example:

For a **35 kDa** protein, the two potential membrane choices are **10 kDa** or **30 kDa** NMWL.

In this case, only the 10 kDa membrane will provide optimal recovery.

Our current product portfolio for the volume range of 0.5ml up to >10L



Microcon	Amicon Ultra 4	Amicon Ultra 15	Centricon Plus-70	Prep/Scale	Pellico
<1 mL	1 – 4 mL	4 – 30 mL	30 – 420 mL	2 – 10 L	>10 L

Pellicon XL

300 mL - 2 L

Stirred Cells

100 - 2000 mL



How to select the best device by volume?



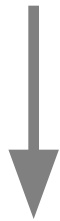
**Maximum
Starting
volume**

< 1 mL

1 to 8 mL

4 – 30 mL

30 - 420mL



**Final
volume**

5 - 15 μ L

50 μ L

200 μ L

350 μ L

Microcon ... the lab standard for small volumes



High recovery design

- Regenerated cellulose membrane
 - low binding
 - highly retentive
 - High recovery
- Invert spin concentrate recovery
- Low binding materials and o-ring seal
- Final conc. Volume 5 - 15 μ L
- cut offs 3K, 10K, 30K, 50K, 100K

Microcon in DNA / RNA applications – Use the Selection Guideline in the protocols

Centrifugation Guidelines for Microcon Devices

Ultracel YM Membrane	Color Code	Membrane NMWL	NCO		Maximum g-Force	Spin Times* 25 °C**
			SS	DS		
YM-3	Yellow	3,000	10	10	14,000	100
YM-10	Green	10,000	30	20	14,000	30
YM-30	Clear	30,000	60	50	14,000	12
YM-50	Rose	50,000	125	100	14,000	12
YM-100	Blue	100,000	300	125	14,000 500 ¹	12

NMWL: Nominal molecular weight limit in Daltons (proteins)

NCO: Nucleotide cut-off for RNA/DNA (SS = single stranded, DS = double stranded)

*Time in minutes; 500 µL samples reduced to 10 µL.

**Sample concentration using Microcon devices at 4 °C typically takes twice as long as that at 25 °C.

¹For RNA/DNA samples using Microcon devices with Ultracel YM-100 membrane, the recommended g-force is 500 × g.

NOTE: DNA sample concentration using Microcon devices generally requires shorter spin times. For example, at 25 °C, a 10 µL final volume of an RNA/DNA sample can be achieved in a Microcon device with Ultracel YM-30 membrane in just 8 minutes.

The next stage in centrifugal filter devices.



1 – 4 mL

4 – 15 mL

Amicon® Ultra



Designed for speed and convenience -- without sacrificing recovery

- 5 - 45 minute processing times
- Single-spin operation -- recover concentrate by pipette
- Universal rotor compatibility

Low-binding Ultracel ultrafiltration membrane

- Retentate recoveries >90%
- Broad chemical compatibility

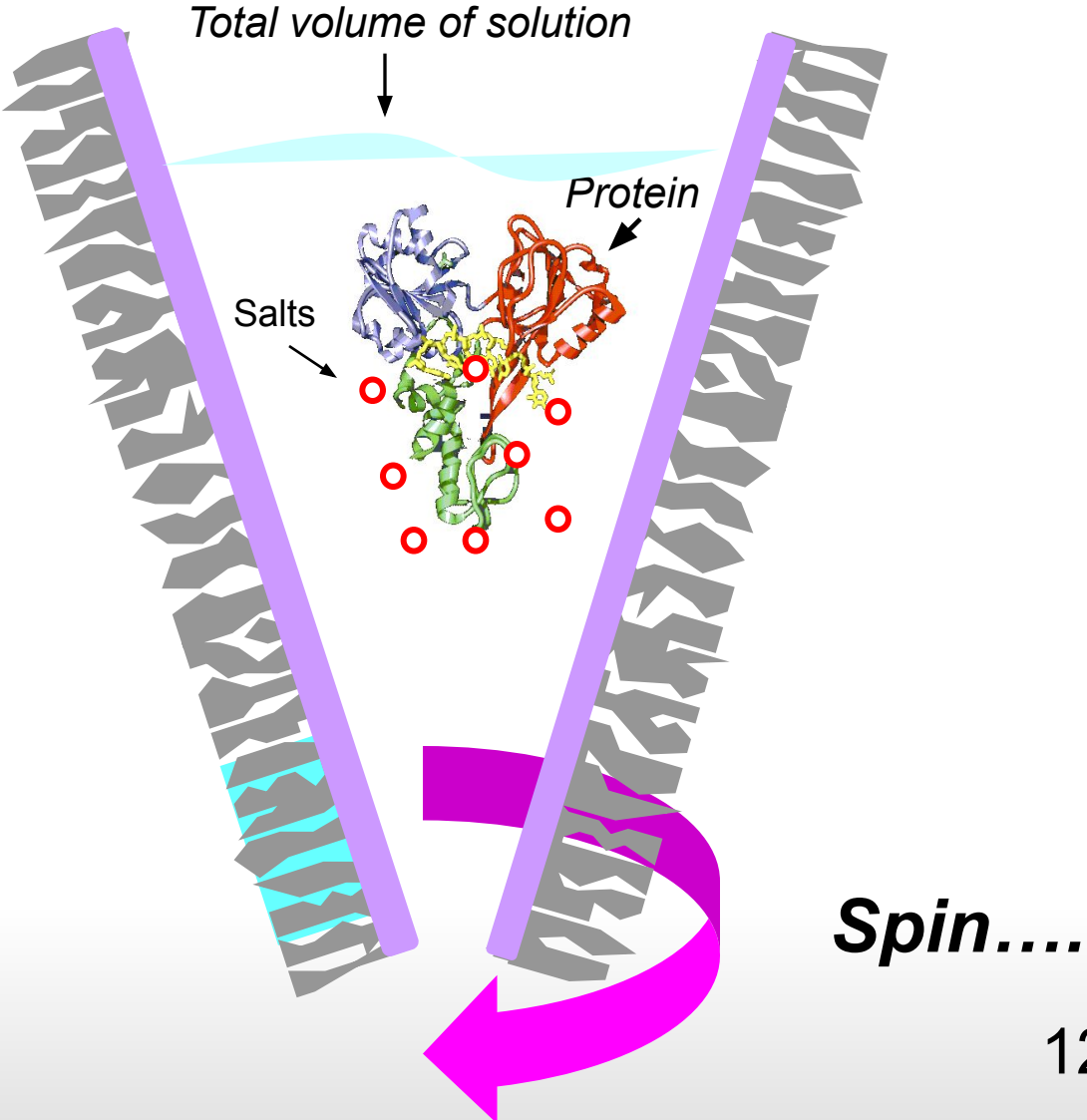
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Selecting a Centrifugal Device

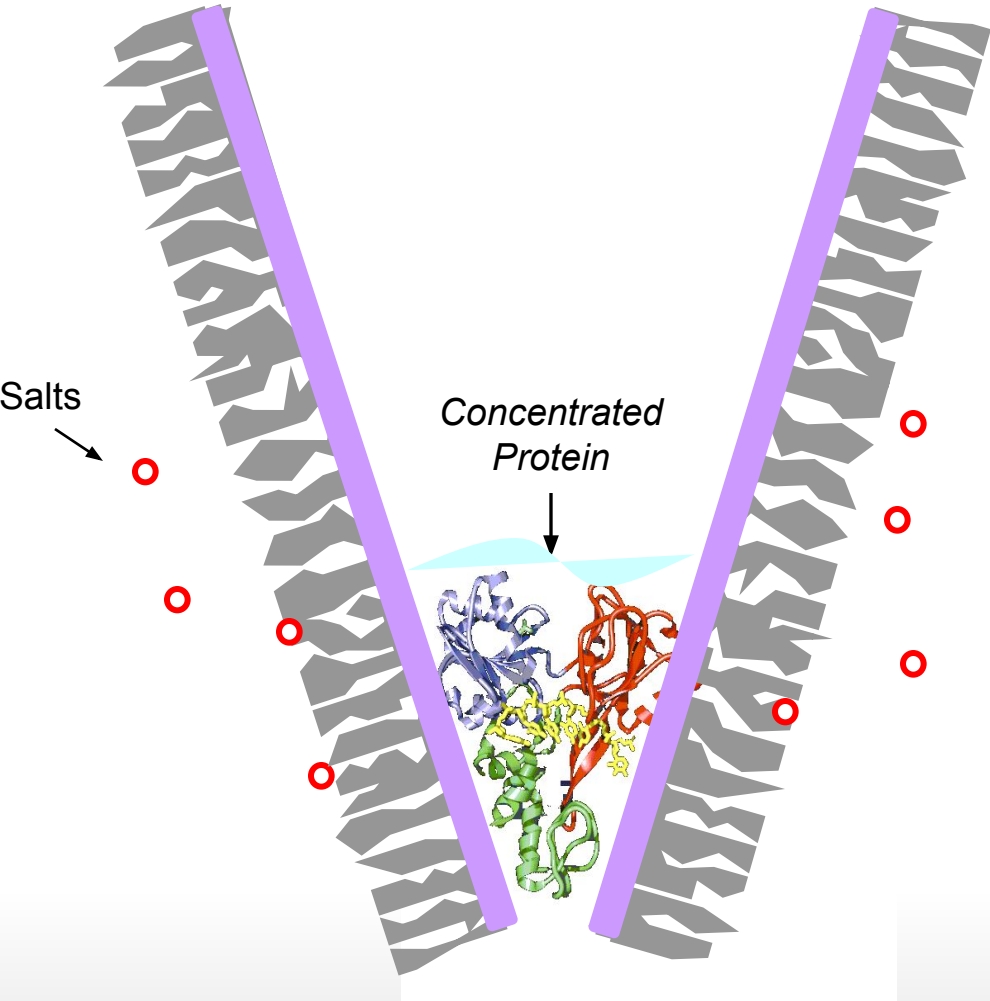
- A vertical membrane design applies the principles of TFF and accelerates concentration times
- Throughput is important when working with larger volumes.
- A dead-stop can prevent spinning to dryness



Amicon Ultra Mode of Action



Amicon Ultra Mode of Action



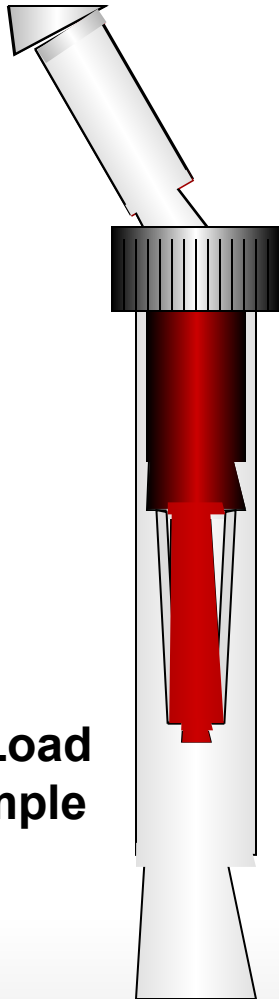
1 – 4 mL

4 – 15 mL

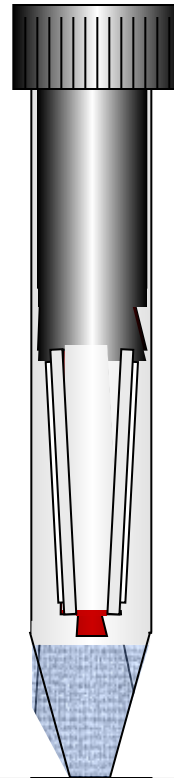
Fast and Easy

Single-spin operation

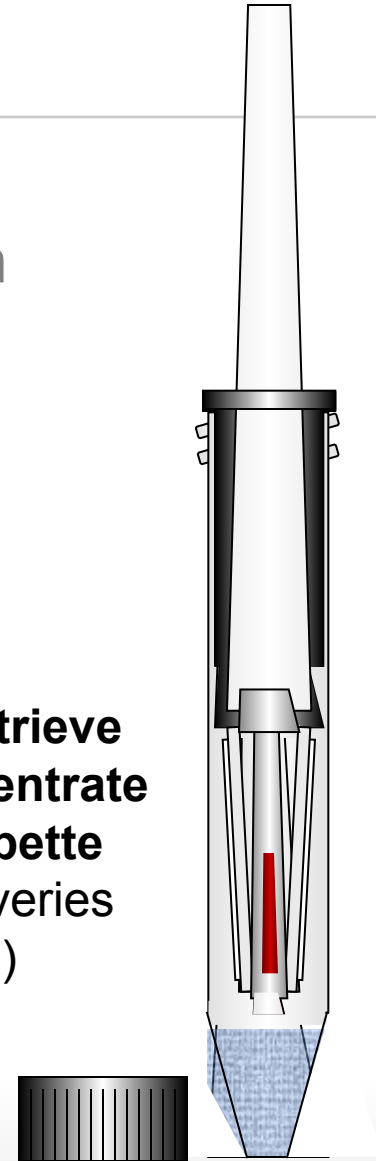
1) **Load sample**



2) **Spin**
(typical spin times 10-20 minutes)



3) **Retrieve concentrate**
by pipette (recoveries >90%)



Select the correct device size:

Samples **2 - 8 mL** = **4 mL** device
Cut offs: 5K, 10K, 30K, 100K



Samples **10 - 30 mL** = **15 mL** device
Cut offs: 5K, 10K, 30K, 100K



Amicon Ultra - Product Offering



Model & Volume	Membrane Type	MWCO	8/pk	24/pk	96/pk
Amicon Ultra-4 ml	Ultrace low-binding cellulose	3 kD	UFC80030	UFC80032	UFC80039
		10 kD	4	4	6
		30 kD	UFC80100	UFC80102	UFC80109
		50 kD	8	4	6
		100 kD	UFC80500	UFC80502	UFC80509
			8	4	6
			8	4	6
Amicon Ultra-15 ml	Ultrace low-binding regenerated cellulose	3 kD	UFC90030	UFC90032	UFC90039
		10 kD	8	4	6
		30 kD	UFC90010	UFC90102	UFC90109
		50 kD	8	4	6
		100 kD	UFC90500	UFC90502	UFC90309
			8	4	6
			8	4	6

Centricon Plus-70 Large Volume Device

30 - 70 mL

**Concentrates down to 350 μ L
in 15 to 20 minutes.**

- Efficiency with double core vertical membrane
- Dead stop with invert spin
- Low-binding Ultracel ultrafiltration membrane



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How to select the type of rotor and G force

RPM



RCF (g) max
with each device



Radius of rotation (mm)

g-Force to RPM

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Fixed Angle

Swinging Bucket

RPM **RCF (g)** **Radius of Rotation (mm)**

Microcentrifuge load angle

Amicon Ultra swinging bucket

Centrifuge Plus-70 swinging bucket

1 Measure the Radius of the Rotor
Obtain the radius (r) of the rotor from the manufacturer's specifications or measure the radius as shown above.

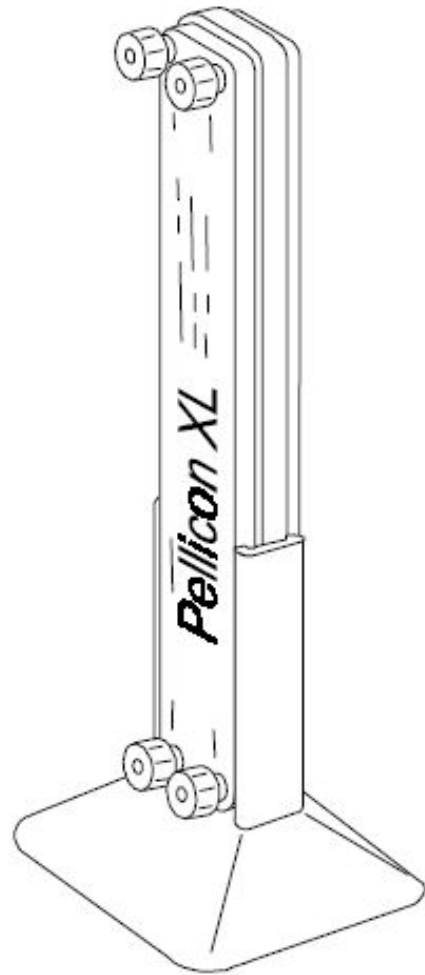
2 Use Nomogram to Calculate RPM
Plot the radius of the rotor and the recommended RCF (g force) of the device on the nomogram and then draw the line to determine RPM. The nomogram is based on the formula of right.

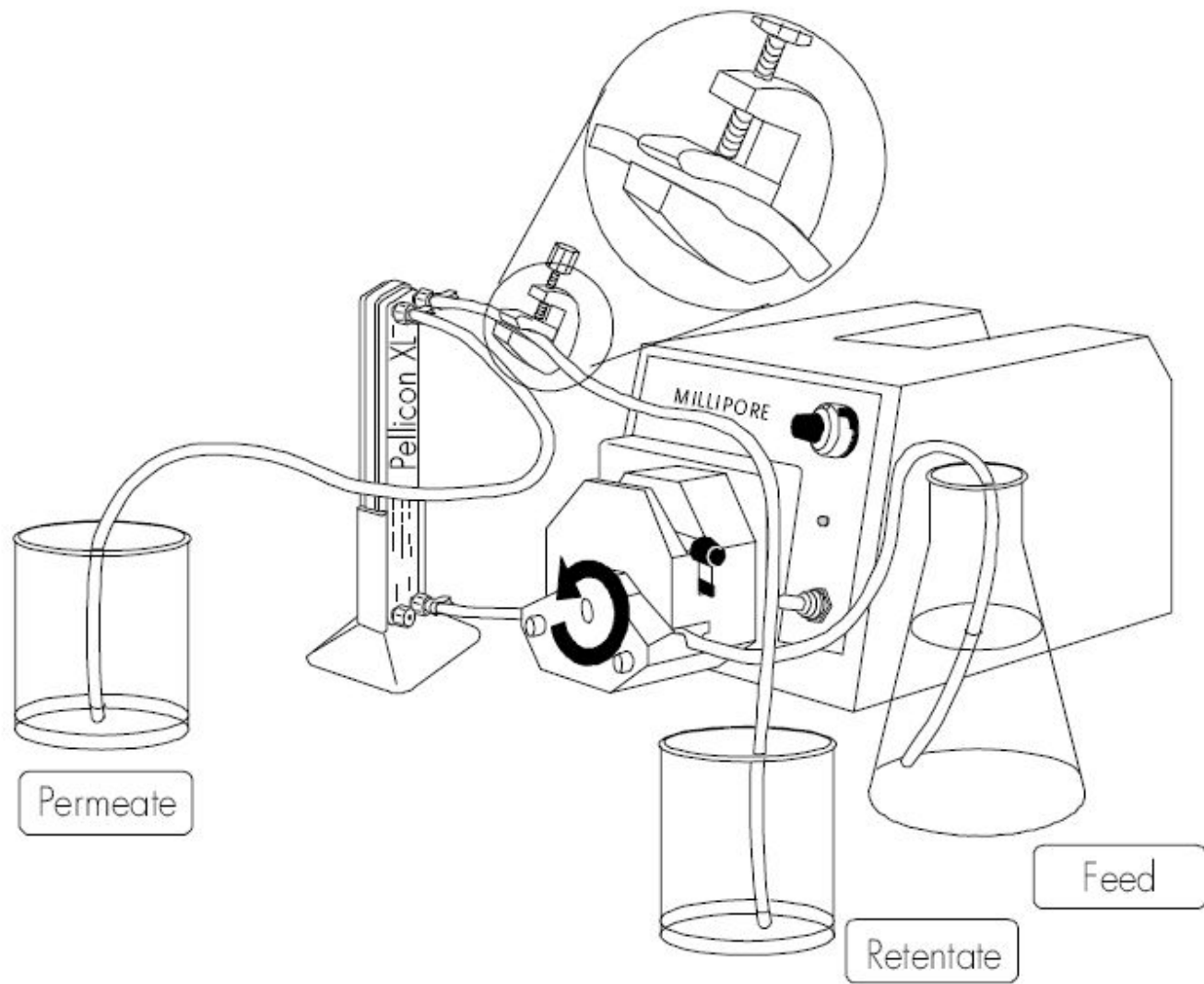
$$RCF = \frac{RPM^2}{(1.118 \times 10^4) \times (\text{Radius in mm})} = RPM$$

AMICON High Speed, High Recovery Ultrafiltration Devices

Microcentrifuge Filter < 1 mL Amicon Ultra-15 Centrifuge Filter < 4 mL Amicon Ultra-15 Centrifuge Filter < 15 mL Centrifuge Plus-70 Centrifuge Filter < 70 mL Amicon XL 50 Cassette < 2 L Steroid Cell < 2 L

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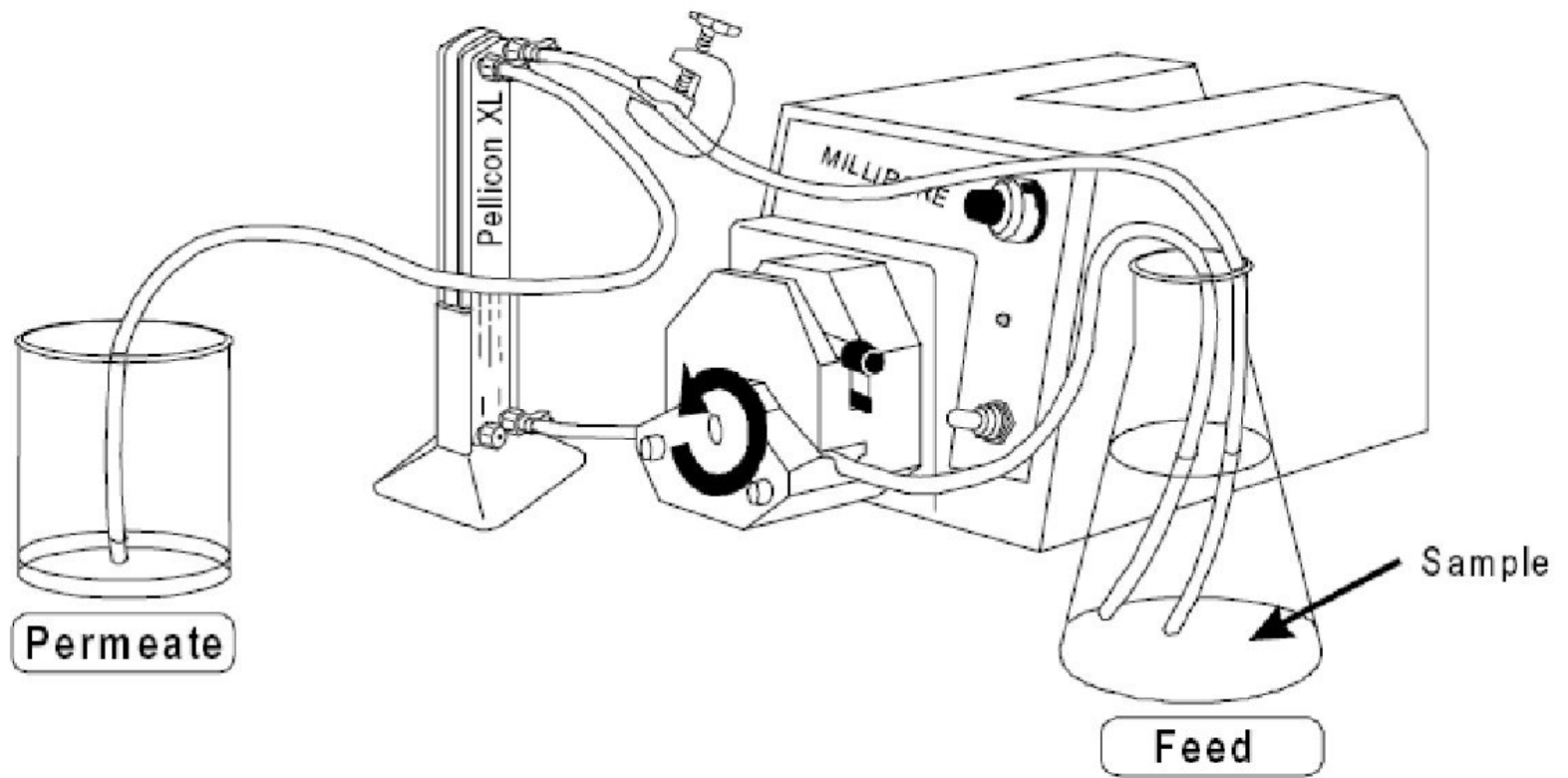


Figure 5: Sample Processing

Section VII: Cleaning

Recommended cleaning solutions (marked by X):

Cleaning Agents	Concentration	Catalogue Numbers with prefix PXB Biomax™ Membrane (polyethersulphone)	Catalogue Numbers with prefix PXC Ultracel™ PLC Membrane (composite regenerated cellulose)	Catalogue Numbers with prefix PXV, PXG, PXH, PXD Durapore® Membrane (PVDF)	Temp °C	pH	Time (min)
NaOH	0.1-0.5N	X			40-45	13-13.7	30-60
NaOH	0.1N		X		25-40	13	30-60
NaOCl	250 ppm	X		X	40-45	10-11	30-60
Triton®-X 100	0.10%	X	X	X	40-45	5-8	30-60
SDS	0.10%	X	X	X	40-45	5-8	30-60
Tween® 80	0.10%	X	X	X	40-45	5-8	30-60
Tergo-zyme® Detergent	0.20%	X	X	X	40-45	9-10	30-60

Do not exceed 30 psi inlet for cleaning cycle.

Section VIII: Storage

Recommended storage solutions:

Storage solution	Concentration	Catalogue Numbers with prefix PXB Biomax Membrane (polyether-sulphone)	Catalogue Numbers with prefix PXC Ultracel PLC Membrane (composite regenerated cellulose)	Catalogue Numbers with prefix PXV, PXG, PXH, PXD Durapore Membrane (PVDF)
NaOH	0.1N	X		
NaOH	0.05N		X	
Lysol® (BAK)	0.1%	X	X	X
Sodium Azide	0.05%	X	X	X
H ₃ PO ₄	0.1N	X	X	X

Centrifugal devices and more....

[Application Information available](#)

Amicon Bioseparations Information Topics

- Detergent removal
- Passivation (reduce protein binding)
- Desalting and buffer exchange
- Depyrogenation
- Protein, DNA, RNA purification
- Protein and DNA concentration
- PCR product purification
- Extraction DNA from gels
- Peptide Concentration
- UF Selection guide

Thank for your attention



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You got into science to
answer the big questions.
Don't let the little ones
slow you down.