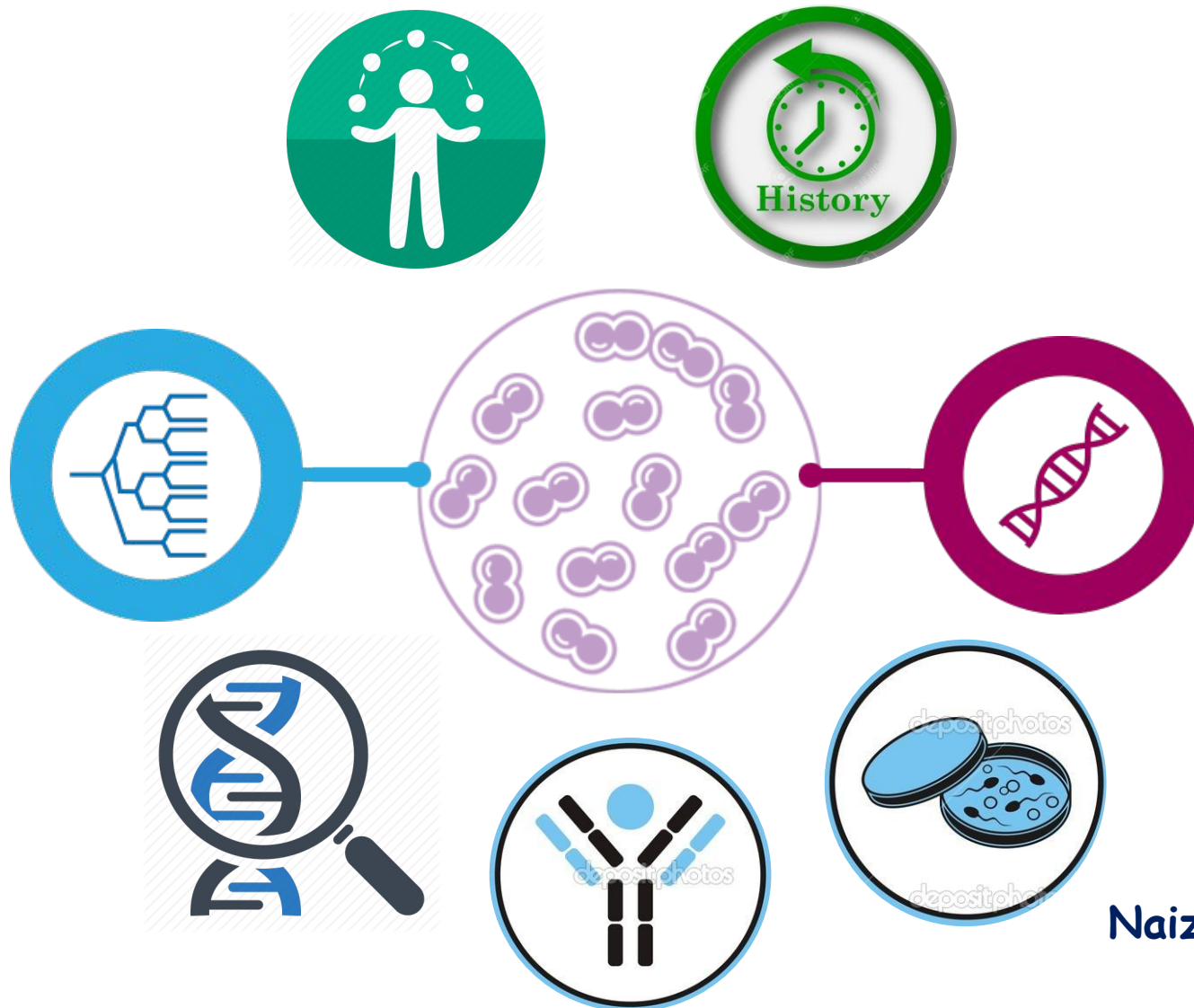


Lactococcus sp.



Done by:
Naizabayeva D.
BT 16-02

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4. Biology (Morphological, physiological and biochemical features)
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Introduction

Lactic acid bacteria are among the most important groups of microorganisms used in food fermentations. They contribute to the taste and texture of fermented products and **inhibit food spoilage** bacteria by producing growth-inhibiting substances and large amounts of lactic acid. As agents of fermentation LAB are involved in making **yogurt, cheese, cultured butter, sour cream, sausage, cucumber pickles, olives and sauerkraut**, but some species may spoil beer, wine and processed meats.

Meanwhile, they have a great application in sphere of pharmacy, due to their antimicrobial ability and safety for the human, taken as probiotics, produce practically important bactericines, and recently practiced as a delivery agent for therapeutics and vaccines.

2. History and discovery

The common organism associated with the souring of milk was first described by **Lister (1878)** as '**Bacterium lactis**'. In the next thirty years a variety of names and descriptions, including **Str. lacticus** of **Kruse (1903)**, were added to the literature, and it was left to **Lohnis (1909)** to clarify the situation. He agreed that it should be placed in the genus **Streptococcus** and suggested that it should be named '**Streptococcus lactis**', thus keeping **Lister's** original species name. The various synonyms encountered are very numerous and were summarized by **Breed (1928)**. **Bergey (1939)** has adopted this list in its entirety. **Orla Jensen's (1919)** description of **Str. lactis** is as follows. The optimum temperature for growth is 30°C., and when freshly isolated it will coagulate sterile milk at this temperature in less than 24 hr. At the optimum temperature it occurs as a diplococcus or as short chains. Growth below 10 °C. and above 40°C is in general poor. **Str. lactis** is characterized by failure to ferment **sucrose**.

2. History and discovery

This group of bacteria, previously designated the lactic streptococci (*Streptococcus lactis* subsp. *lactis* or *S. lactis* subsp. *cremoris*) was placed in this new taxon in **1985 by Schleifer**. This discovery was investigated according to results of **nucleic acid hybridization studies** and **immunological relationships of superoxide dismutase**, which in its turn demonstrated that *Streptococcus lactis* (and its subspecies), *Lactobacillus xylosus*, *Lactobacillus hordniae*, *S. garvieae*, *S. plantarum* and *S. raffinolactis* are closely related to each other but not to other streptococci. Therefore it was proposed that these taxa be transferred to a new genus *Lactococcus* gen.nov. as *Lactococcus lactis* subsp. *lactis* (including former *S. lactis* subsp. *diacetylactis* and *Lactobacillus xylosus*) comb.nov., *L. lactis* subsp. *cremoris* comb.nov., *L. lactis* subsp. *hordniae* comb.nov., *L. garvieae* comb.nov., *L. plantarum* comb.nov. and *L. raffinolactis* comb.nov. The relatedness of these organisms has also been demonstrated by the similarity of their lipoteichoic acid structures, lipid pattern, fatty acid and menaquinone compositions.

2. History and discovery



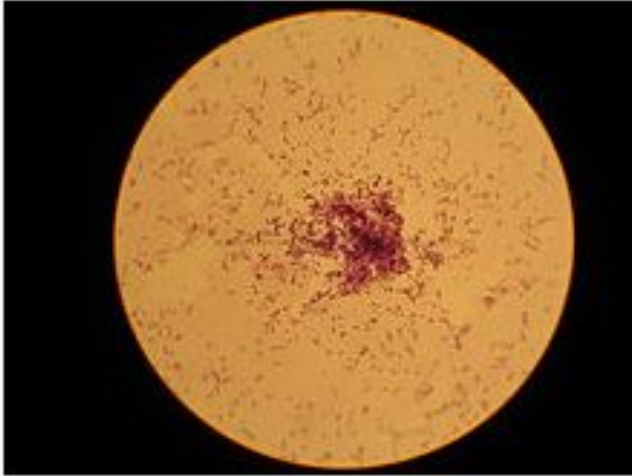
Table 1 Comparison of suggested groupings for streptococci and enterococci

Sherman 1937	Jones 1978	Bridge and Sneath 1983	Bergey 1986	Schleifer and Kilpper-Bälz 1987
Pyogenic	Pyogenic	Pyogenic Parapyogenic	Pyogenic	Pyogenic
	Pneumococci	Pneumococcal		<i>Strep. oralis</i> group (including ' <i>Strep. milleri</i> ') <i>Strep. mutans</i> group
Viridans	Oral	Viridans Paraviridans	Oral	
	Other streptococci Anaerobic streptococci	Thermophilic	Other streptococci Anaerobic	Other streptococci Not streptococci
<i>Enterococcus</i>	Faecal	Enterococcal	Enterococci	<i>Enterococcus</i> (separate genus)
Lactic	Lactic	Lactic	Lactic acid cocci	<i>Lactococcus</i> (separate genus)



3. Taxonomy

Lactococcus



Lactococcus lactis

Scientific classification

Kingdom: Bacteria
Division: Firmicutes
Class: Bacilli
Order: Lactobacillales
Family: Streptococcaceae
Genus: ***Lactococcus***
Schleifer et al. 1986

Species

- *L. chungangensis*
- *L. formosensis*
- *L. fujiensis*
- *L. garvieae*
- *L. lactis*
 - *L. lactis* subsp. *cremoris*
 - *L. lactis* subsp. *hordniae*
 - *L. lactis* subsp. *lactis*
 - *L. lactis* subsp. *tructae*
- *L. piscium*
- *L. plantarum*
- *L. raffinolactis*
- *L. taiwanensis*

4. Biological features: morphological

Lactococci are homofermentative, **microaerophilic** **Gram-positive** bacteria characterized by ovoid cells **0,5-1,2x0,5-1,5 mkrm** in size, which appear individually, in pairs, or in chains. Non-motile, do not form spores or capsules.



4. Biological features: physiological

physiology

Genus	<i>Lacto- bacillus</i>	<i>Entero- coccus</i>	<i>Lacto- coccus</i>	<i>Leuconostoc</i>	<i>Pedio- coccus</i>	<i>Strepto- coccus</i>
Characteristic						
Morphology	rods	cocci	cocci	cocci	cocci in tetrads	cocci
CO ₂ from glucose*	±	-	-	+	-	-
Growth						
at 10°C	±	+	+	+	±	-
at 45°C	±	+	-	-	±	±
in 6.5% NaCl	±	+	-	±	±	-
at pH 4.4	±	+	±	±	+	-
at pH 9.6	-	+	-	-	-	-
Lactic acid configuration	D, L, DL	L	L	D	L, DL	L

+ positive; - negative; ± varies between species

* test for homo- or heterofermentation of glucose: - homofermentation

+ heterofermentation

4. Biological features: physiological

Protein metabolism

Nitrogen source -> 1. free amino acids in the composition of milk
2. Casein, which composes 80% of all proteins present in milk

1. Essential amino acid for most Lactococci are **isoleucine, leucine, valine, histidine, methionine**. The concentration of these amino acids in milk is less than **2 mg/l**. Amount of nitrogen in this case provides only **2%** of the final cell density.

2. Casein, becomes the primary nitrogen source after nonprotein nitrogen is depleted.

The enzymes that form proteolytic system - a cell wall-associated proteinase, an extracellular peptidase (s), amino acid transport system, peptide transport system and intracellular peptidases.

The key enzyme - is a cell-wall associated proteinase (PI- or PIII- type proteinase [PrtP])

Transport systems- di- and tripeptide and an oligopeptide transport system.

4. Biological features: physiological

Characteristic	1	2	3	4	5	6	7	8
Growth at:								
4 °C	+	-	+	-	-	-	-	-
40 °C	-	+	-	-	-	-	-	+
Growth in 4 % NaCl	-	+	-	+	-	-	-	+
Ammonia from arginine	-	+	-	-	-	-	+	+
Acid from:								
Amygdalin	+	+	+	+	-	-	-	+
Galactose	-	+	+	+	w	+	-	+
Lactose	-	-	+	-	-	+	-	+
Maltose	+	+	+	+	+	+	-	+
Melibiose	-	-	+	-	+	+	-	-
Melezitose	-	-	+	+	-	-	-	-
Methyl α -D-glucoside	-	-	+	+	-	-	-	w
Methyl α -D-mannoside	-	-	+	-	-	-	-	-
Raffinose	-	-	+	-	+	+	-	-
Sucrose	+	-	+	+	+	+	+	-
Trehalose	w	+	+	+	+	+	+	+
Turanose	w	-	+	+	-	-	-	-
D-Xylose	-	-	+	-	+	+	-	+
Hydrolysis of aesculin	+	+	+	+	-	-	-	-

Strains: 1, CAU 28T;

2, *L. garvieae* KCTC 3772T;

3, *L. piscium* DSM 6634T;

4, *L. plantarum* DSM 20686T;

5, *L. raffinolactis* DSM 20443T;

6, *L. lactis* subsp. *cremoris* KCCM 40699T;

7, *L. lactis* subsp. *hordniae*

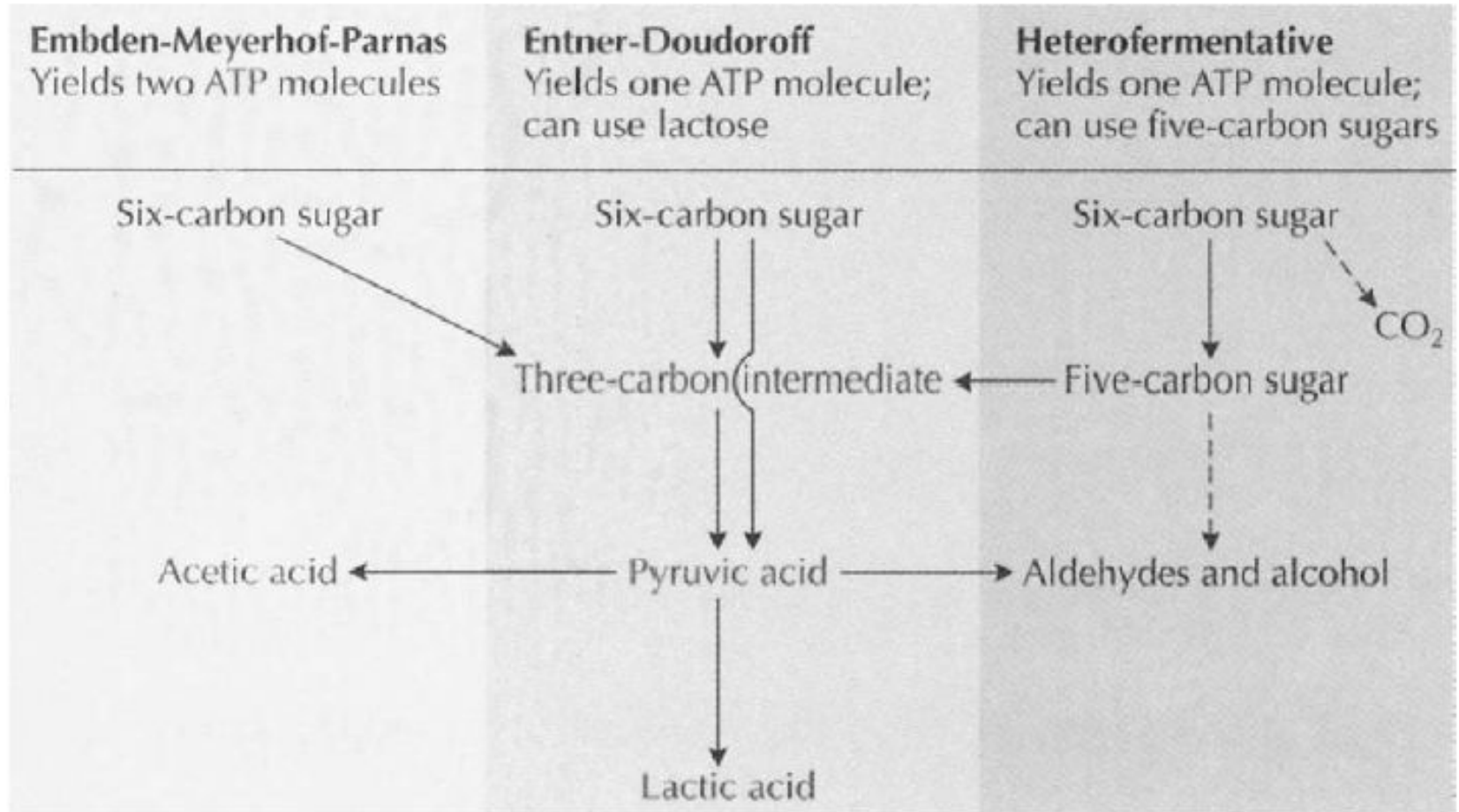
KCTC 3768T;

8, *L. lactis* subsp. *lactis* KCTC 3769T.

(+) Positive, (-) negative, w- weakly positive

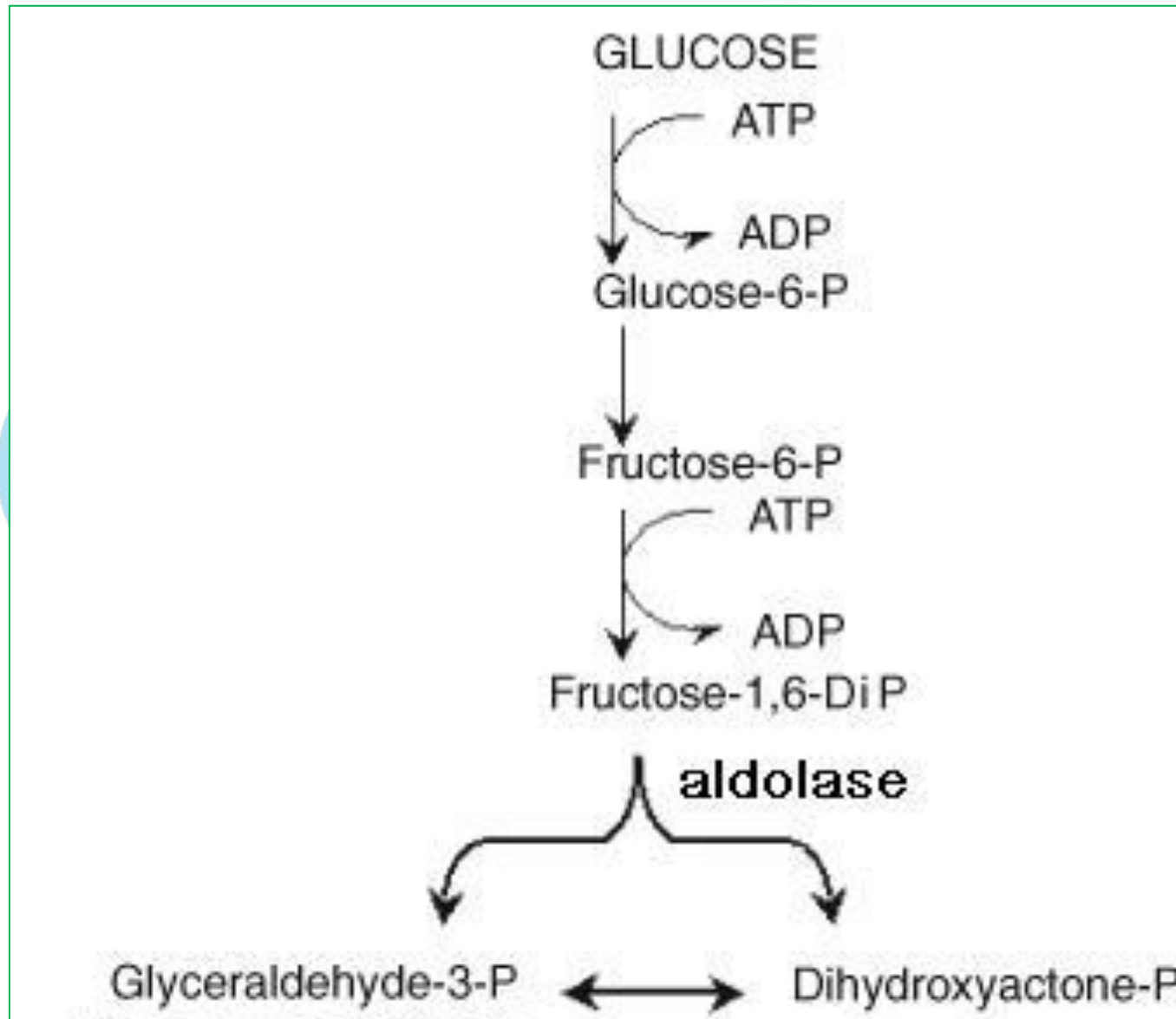
4. Biological features: biochemical

Figure 18.2 Simplified catabolic pathways used in fermented foods.

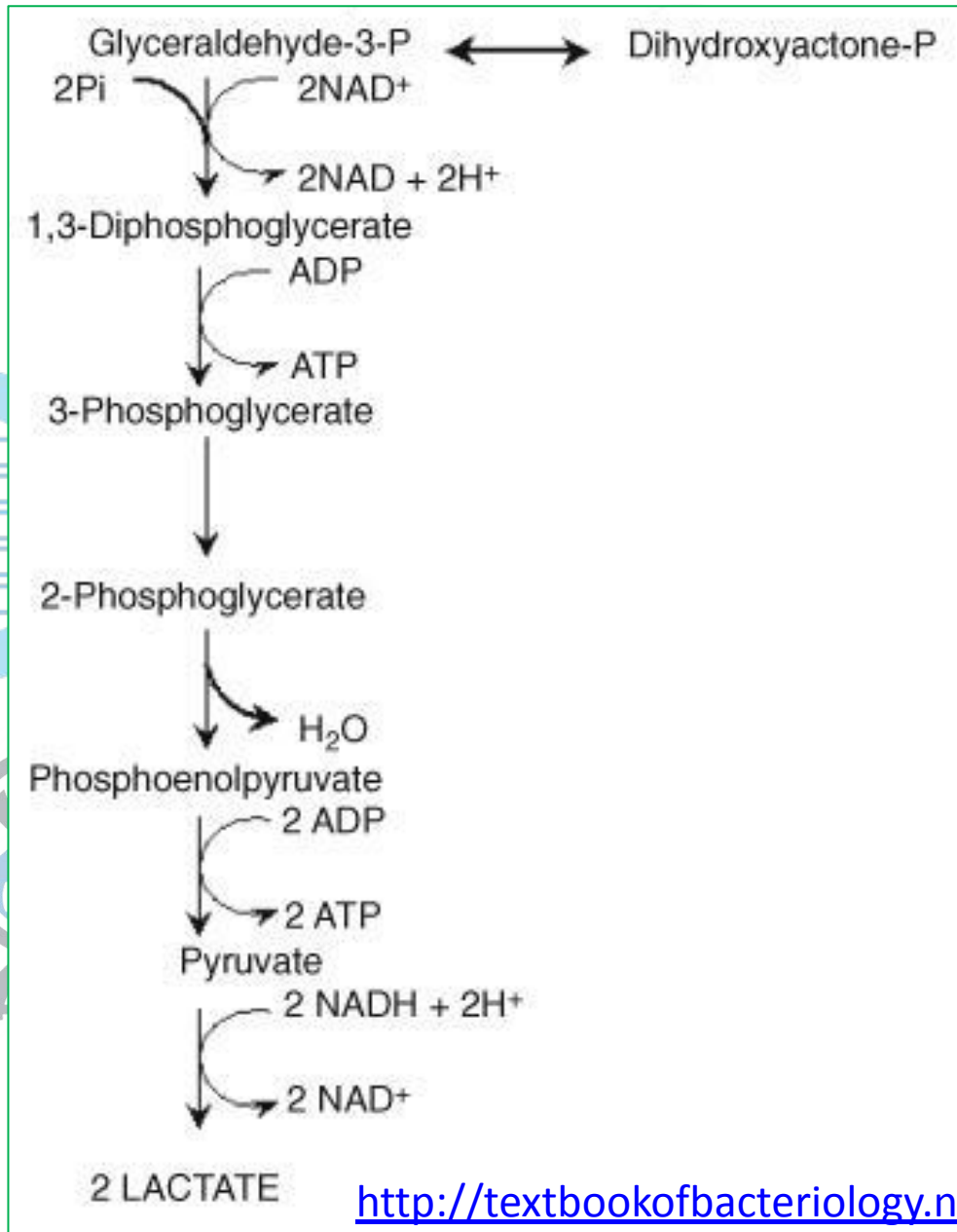


Lactococcus sp. classified as **homofermentative** or **homolactic** , cause the end product is only lactic acid and correspond to **Embden-Meyerhof-Parnas** catabolic pathway. [Montville et al. **FOOD MICROBIOLOGY/Chapter 18**]

4. Biological features: biochemical



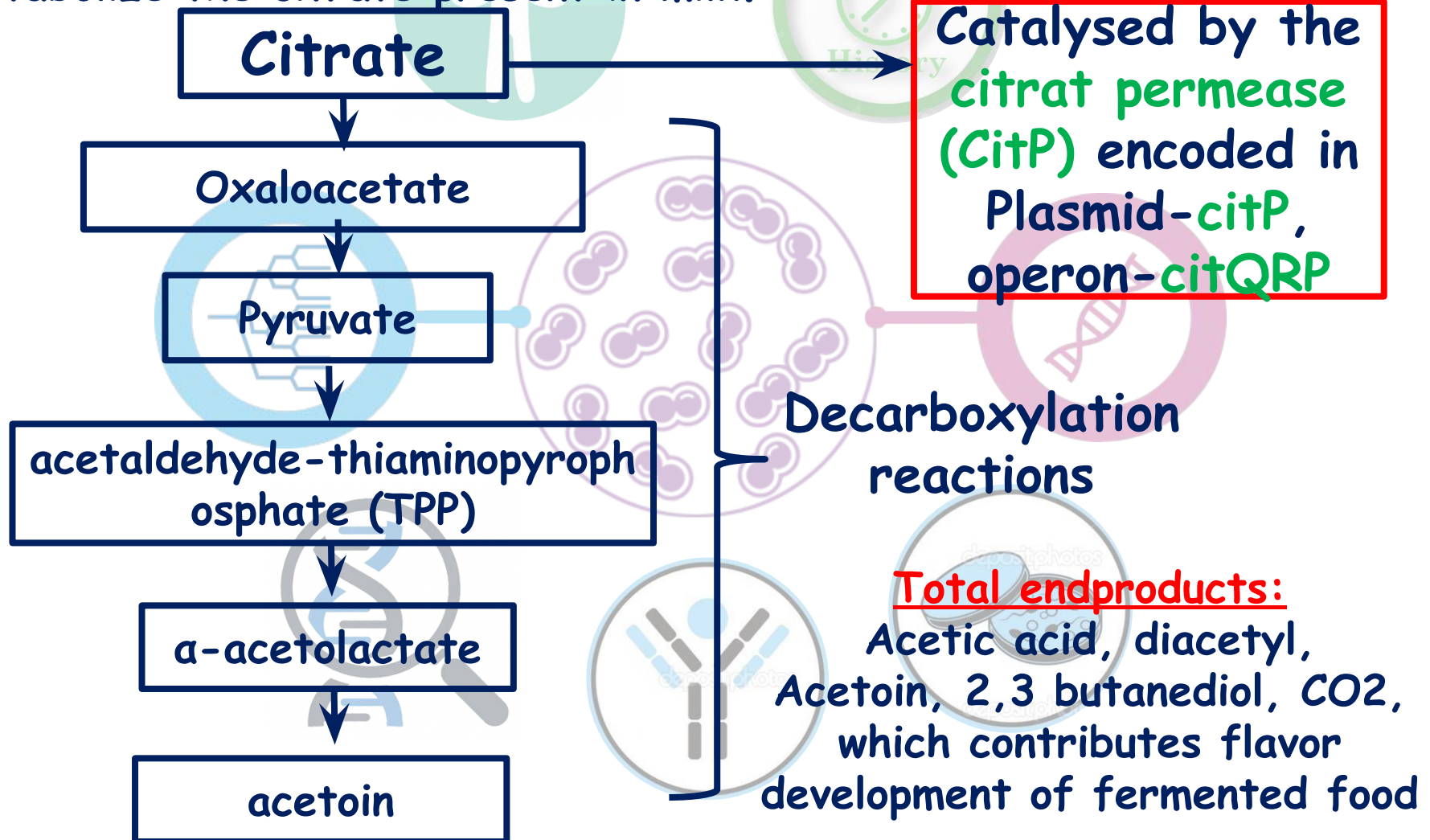
4. Biological features: biochemical



4, Biological features: biochemical

Capability to metabolize citrate

Only *Lc. lactis* subsp. *lactis biovar diacetylactis* has the ability to metabolize the citrate present in milk.



Detection and analysis

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graph TD; A[Detection and analysis] --> B[1. Morphological]; A --> C[2. Physiological / biochemical]; A --> D[3. Serological]; A --> E[4. Genetic]; B --> B1[Microscopy (Gram staining)]; C --> C1[Medium associated tests (antibiotic resistance, carbohydrate fermentation, acetoin production and etc.), API 50CHL strips (fermentation of 49 carbohydrates and esculin hydrolysis), Biolog plate (96 carbohydrates)]; D --> D1[Slide agglutination, fluorescent antibody staining, flow-cytometry based methods]; E --> E1[PCR-based identification]; E --> E2[16S rRNA sequencing]; E1 --> E2;
```

The diagram illustrates a flowchart for the detection and analysis of bacteria. It starts with a central box labeled 'Detection and analysis' in red. From this box, four arrows point to different categories of tests: 1. Morphological (blue), 2. Physiological / biochemical (green), 3. Serological (yellow), and 4. Genetic (purple). Each category then leads to specific methods: Morphological leads to Microscopy (Gram staining); Physiological / biochemical leads to Medium associated tests (antibiotic resistance, carbohydrate fermentation, acetoin production and etc.), API 50CHL strips (fermentation of 49 carbohydrates and esculin hydrolysis), and Biolog plate (96 carbohydrates); Serological leads to Slide agglutination, fluorescent antibody staining, and flow-cytometry based methods; Genetic leads to PCR-based identification and 16S rRNA sequencing, with an arrow indicating that PCR-based identification leads to 16S rRNA sequencing. The background features various circular icons related to microbiology, including a person with a network, a clock labeled 'History', a DNA helix, and petri dishes.

1. Morphological

Microscopy
(Gram staining)

3. Serological

Slide agglutination,
fluorescent
antibody staining,
flow-cytometry
based methods

2. Physiological / biochemical

Medium associated tests
(antibiotic resistance,
carbohydrate
fermentation, acetoin
production and etc.),
API 50CHL strips
(fermentation of 49
carbohydrates and
esculin hydrolysis), Biolog
plate (96 carbohydrates)

4. Genetic

PCR-based
identification

16S rRNA
sequencing

5. Phenotypic analysis: morphological

1. Cultural characteristics (color, margine, transparence of colonies and etc.)
2. Microscopic observation (Gram straining)

BUT, It often happens that cells of **lactococci** themselves extend into a chain, which makes them difficult to differentiate from **lactobacilli**. The group consisting of **Streptococcus**, **Enterococcus** and **Leuconostoc** also forms cocci that occur as chains or pairs, so it is difficult to distinguish these genera from **Lactococcus** genera on a morphological basis.

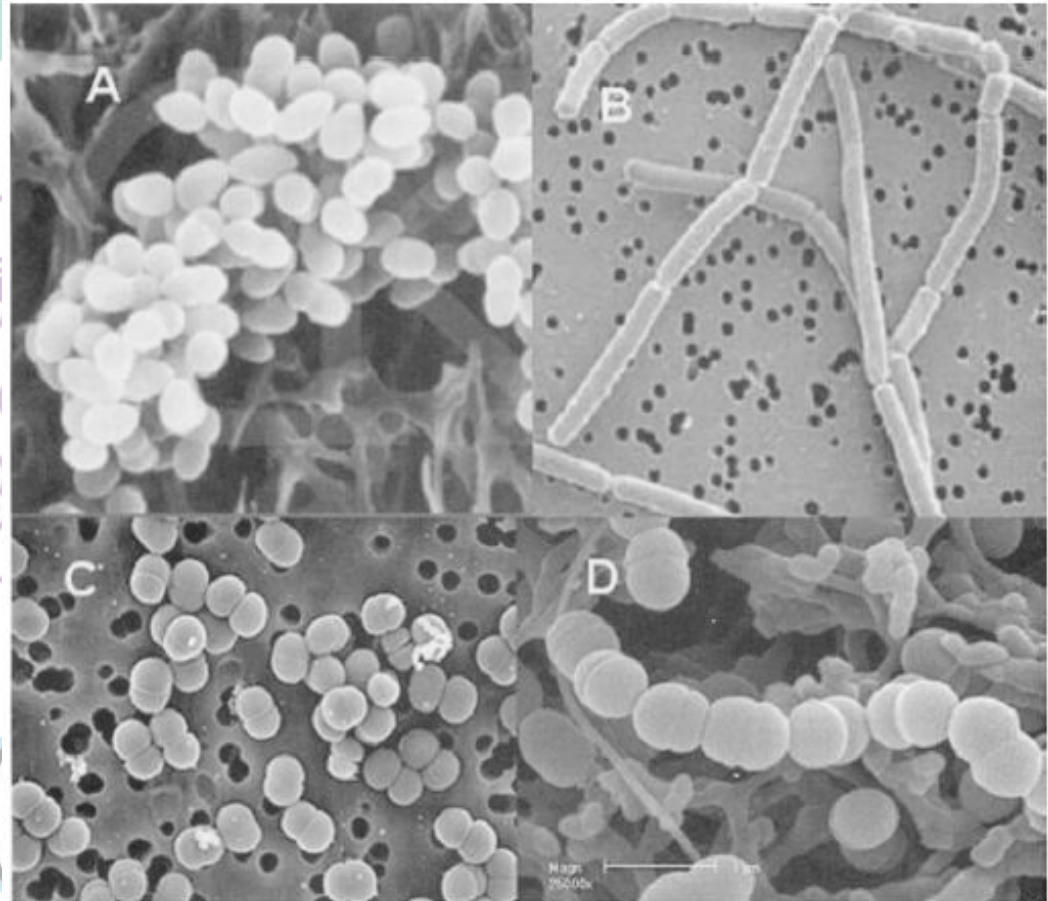


Figure 18.3 LAB associated with food fermentations. (A) *Lactococcus*; (B) *Lactobacillus*; (C) *Pediococcus*; (D) *Leuconostoc*. (B and C) Courtesy of the U.S. Department of Energy Joint Genome Institute. (D) Reprinted from G. Kaletunç, J. Lee, H. Alpas, and F. Bozoglu, *Appl. Environ. Microbiol.* **70**:1116–1122, 2004, with permission from the American Society for Microbiology.

5. Phenotypic analysis: physiological

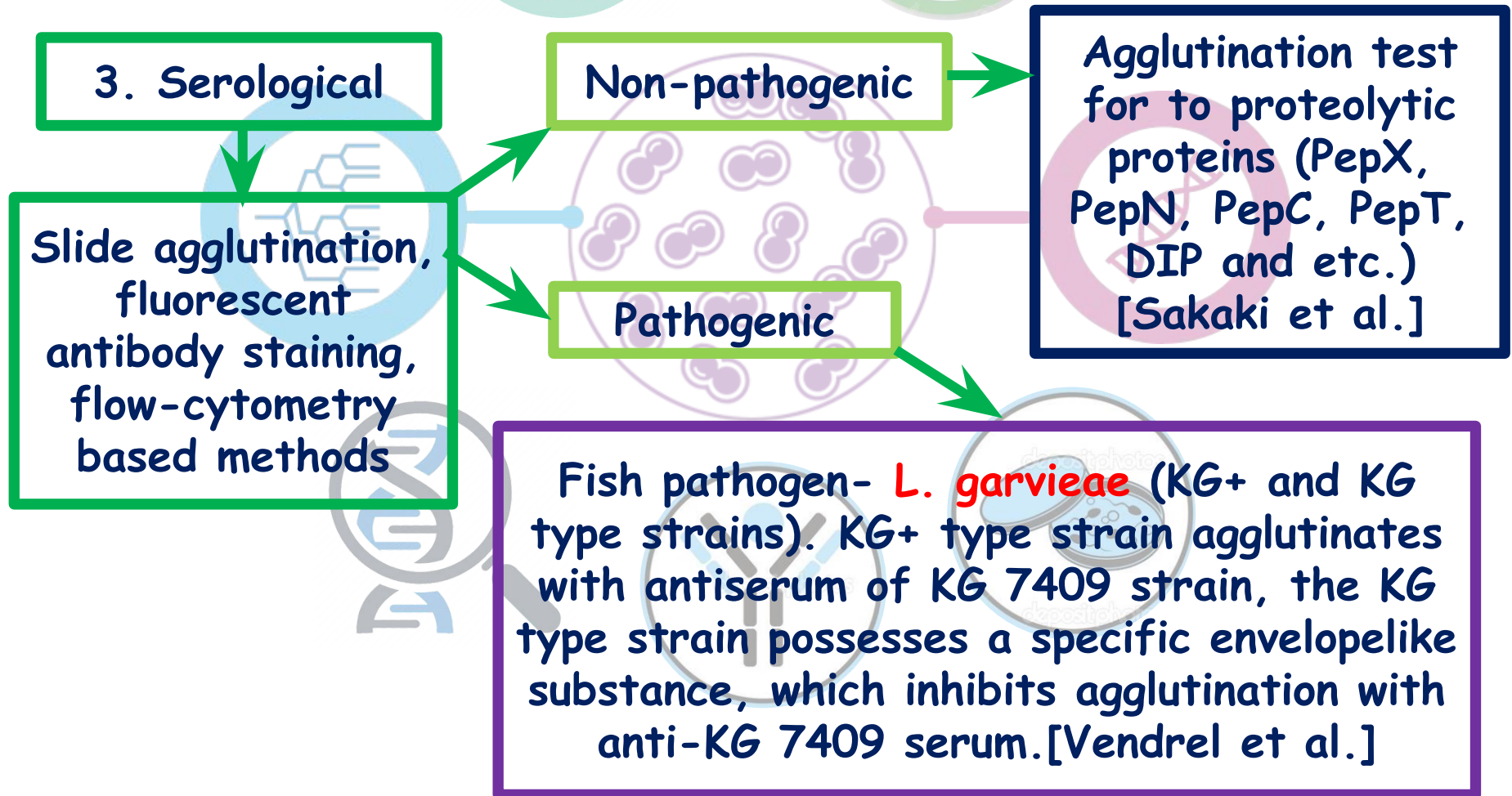
Differentiation of *Lactococcus* species

Species	PYR	VP	Arg	Lac	Man	Mel	Raf	Clind
<i>L. lactis</i> subsp. <i>lactis</i>	v	+	+	+	v	-	-	S
<i>L. lactis</i> subsp. <i>cremoris</i>	-	-	+	+	-	-	-	S
<i>L. lactis</i> subsp. <i>hordiae</i>	-	-	+	-	-	-	-	S
<i>L. garvieae</i>	+	+	+	+	+	-	v	R
<i>L. plantarum</i>	-	-	-	-	+	+	-	
<i>L. raffinolactis</i>	-	-	-	-	v	v	+	
<i>L. xyloso</i>	-	-	+	-	+	-	-	

Acid formation in: **Lac**=lactose, **Man**=mannitol, **Raf**=raffinose, **Mel**-melibiose **Arg**=deamination of arginine, **PYR**=pyrrolidonylarylamidase, and **VP**=Voges-Proskauer* (acetoin detection), **Clind**- clindamycin, (+) = >90% positive, (-) = <10% positive, v = 60-90% strains positive

6. Immunological (serological) analysis

Generally discovery of lactococcus as a separate genera was investigated according to results of **nucleic acid hybridization studies** and **immunological relationships of superoxide dismutase**. According **Lancefield et al.** representatives of **Lactococcus sp.** were classified into serological N group.



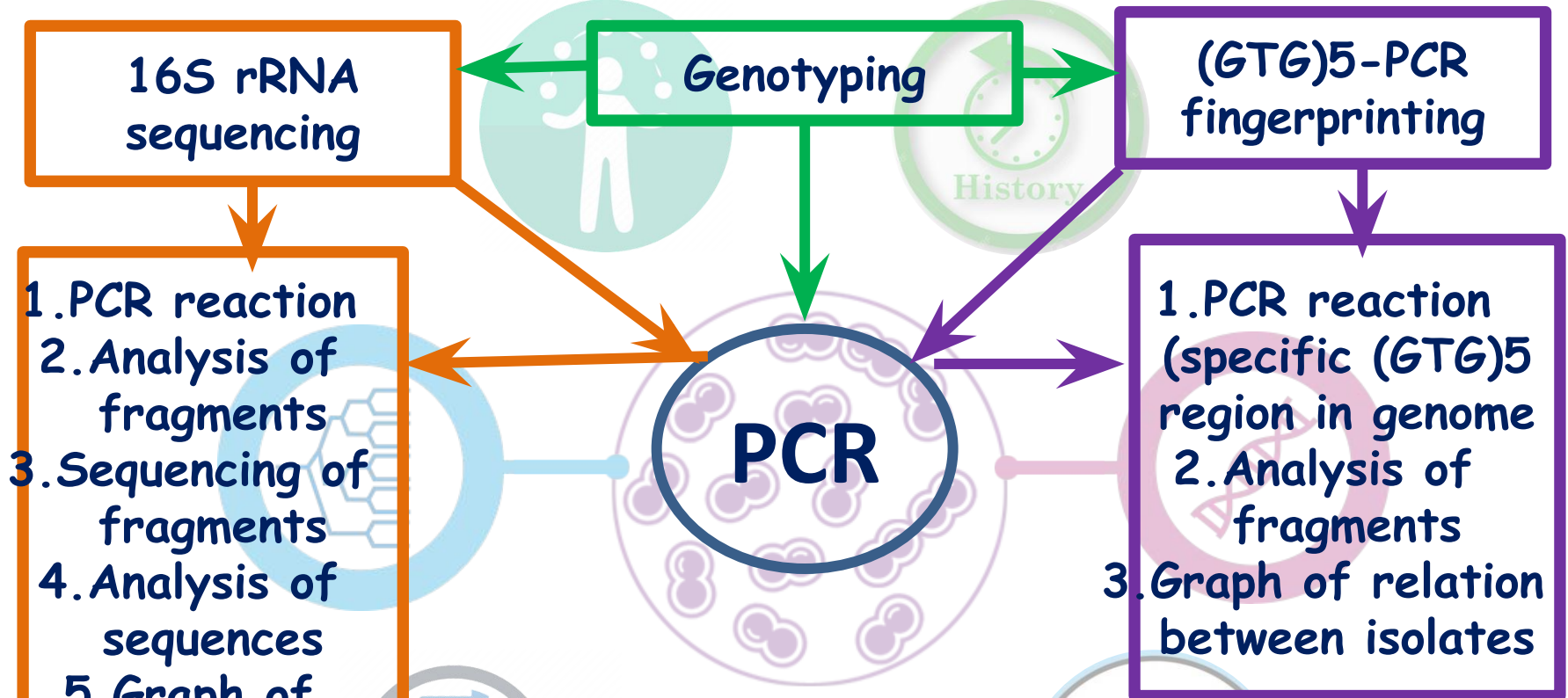
7. Genotypic techniques

Currently, there are two *L. lactis ssp. cremoris* and *L. lactis ssp. lactis*. that have been sequenced for public release. This investigations has a crucial role in understanding and manipulation of fermentation process to obtain desired product , also for fundamental researches of phylogenic diversity and epidemiology of pathogenic strains (*L. garvieae*).

Methods

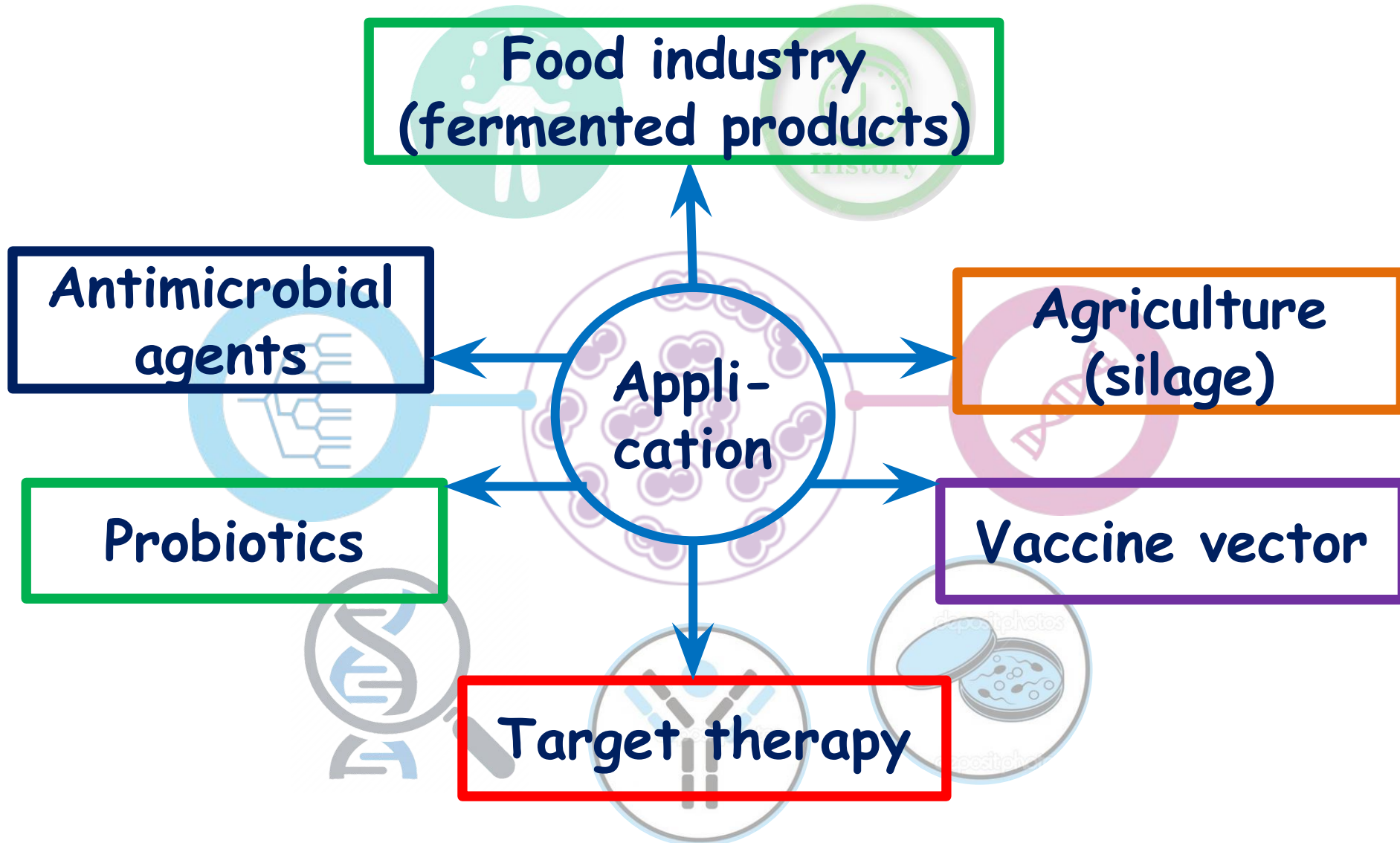
1. 16S rRNA sequencing
2. (GTG)₅-PCR fingerprinting
3. Genotyping IS elements (insertion elements)
4. Randomly amplified polymorphic DNA-polymerase chain reaction (RAPD-PCR)
5. Multilocus sequence typing (for example 7 loci *atpA*, *rpoA*, *pheS*, *pepN*, *bcaT*, *pepX*, and 16S rRNA gene)

7. Genotypic techniques: General scheme



* So general mechanism is the same, the difference is in studying (genotyping) marker, the length and composition of which require designing specific primers and settling the reaction parameter. One more aspect that should be mentioned is the choose of marker for genotyping, cause the discriminatory index has a crucial role in diversification of isolates by strains.

8. Manipulation (genetic or non-genetic)



8. Manipulation (genetic or non-genetic)

1. Food industry: cheese, butter, buttermilk, sour cream and etc (non-genetic/genetic)
2. Agriculture, e.g. silage (non-genetic)
3. Source of antimicrobial agents and preservatives called bacteriacines. For example "Nisin". (non-genetic/ may include genetic manipulation for commercial overproduction)
4. Medicine – probiotics (non-genetic)
5. Medicine delivery factor (genetic [gene therapy])
6. New type of recombinant vaccine vector (genetic).

8. Manipulation (genetic or non-genetic)

The food-grade bacterium *Lactococcus lactis* has been extensively investigated during the last two decades as a delivery vector for **therapeutic proteins**, **DNA** and **vaccine antigens**. The bacterium represents a safe, genetically tractable vector capable of producing heterologous therapeutic proteins at mucosal sites. Contributing this, recombinant *L. lactis* strains have been exploited as agents to treat inflammatory **bowel disease**, **allergy** and **cancer**.

Examples of vaccine delivery in practice – tetanus toxin C, pneumococcal diseases, staphylococcal enterotoxin B, *H. pylori* (HspA gene), and etc.

8. Manipulation (non-genetic)

Fermented dairy products wherein *Lactococcus lactis* is the primary organism involved in manufacture.

Product	Principal acid producers	Secondary microflora
Cheese		
Colby, Cheddar, cottage, cream	<i>Lactococcus lactis</i> ssp. <i>cremoris</i>	None
	<i>Lactococcus lactis</i> ssp. <i>lactis</i>	
Blue	<i>Lactococcus lactis</i> ssp. <i>cremoris</i>	Citrate ⁺ <i>Lactococcus lactis</i> ssp. <i>lactis</i> <i>Penicillium roqueforti</i>
	<i>Lactococcus lactis</i> ssp. <i>lactis</i>	
Fermented milk		
Buttermilk	<i>Lactococcus lactis</i> ssp. <i>cremoris</i>	<i>Leuconostoc</i> spp. Citrate ⁺ <i>Lactococcus lactis</i> ssp. <i>lactis</i>
	<i>Lactococcus lactis</i> ssp. <i>lactis</i>	
Sour cream	<i>Lactococcus lactis</i> ssp. <i>cremoris</i>	None
	<i>Lactococcus lactis</i> ssp. <i>lactis</i>	

8. Manipulation (genetic or non-genetic)

In sphere of Food industry, Agriculture and probiotic production general scheme of manipulation is following:
Isolation -> cultivation-> biomass -> (purification, capsulation in case of probiotics or nisin production)-> treatment.

In case of genetic manipulations, the scheme provided in next order:

1. Isolation (donor of gene)-> cultivation-> DNA extraction-> identifying sequence of desired gene-> construction of vector-> transformation of recipient microorganism.

2. Isolation (recipient of gene)-> cultivation -> transformation->selection-> cultivation-> downstream processes (purification, concentration, capsulation, package)-> treatment.

9. Facts

- . *Lactococcus sp.* strains considered to be safe (GRAS) for human and used in dairy product fermentation.
- . The genus contains strains known to grow at or below 7°C .
- . Today they are used extensively in food fermentations, which represent about 20% of the total economic value of fermented foods produced throughout the world.
- . The *Lactococcus lactis ssp. lactis* genome has 2 365 589 units (bp) of DNA, which contain 2 310 predicted genes. About 64% of the genes have assigned roles in the cell, while 20% match other hypothetical genes with unknown function. Almost 16% of the genes bear no resemblance to genes from other species and are considered to be unique to this bacterium.
- . Before 1985, representative of this genera were classified as *Streptococcus* and *Enterococcus sp.*

CONCLUSION

Description of the genus *Lactococcus* gen.nov.
Lactococcus (lac.to.coc'cus, L.n.lac, lactis milk.,
Gr.n.coccus, a grain or berry, M.L.masc.n. *Lactococcus*
milk coccus).

Spheres or ovoid cells occur singly, in pairs or in chains, and are often elongated in the direction of the chain. Gram-positive. Endospores are not formed. Non-motile. Not β -haemolytic. Facultatively anaerobic, catalase negative. Growth at 10°C but not at 45°C . Usually grows in 4 % (w/v) NaCl with the exception of *L. lactis* subsp. *cremoris* which only tolerates 2 % (w/v) NaCl. Chemoorganotrophs. Metabolism- fermentative. The predominant end product of glucose fermentation is L-lactic acid. Most strains react with group N antisera. Some strains possess low levels of menaquinones.

CONCLUSION

The major glycolipid of all strains is *Glc(a1-2)Glc(a1-3)acyI2-Gro*, a constant minor component is *Glc(a1-2), acyl- 6Glc(a1-3)acyI2Gro*. All strains contain phosphatidylglycerol and cardiolipin. Lipoteichoic acid structure and occurrence of aminophospholipids are species rather than genus-specific. Non-hydroxylated long-chain fatty acids are primarily of the straight-chain saturated and monounsaturated types; some strains produce cyclopropane-ring acids. The major fatty acids are hexadecanoic and *cis- 11,12-octadecenoic acids*; *cis-11,12-methylenoctadecanoic* acid is also present in major amounts in most strains with the exception of *L. lactis subsp. hordniae* and *L. raffinolactis*. The G+C content of the DNA ranges from 34 to 43 mol %.

CONCLUSION

Nucleic acid hybridization and comparative immunological studies demonstrate that members of the genus *Lactococcus* are closely related to each other but not to members of the genus *Streptococcus* or *Enterococcus*. Lactococci can be distinguished from streptococci and enterococci by their ability to grow at 10°C but not at 45°C . They are not β -haemolytic; some strains show a weak α -haemolytic reaction. They are non-motile.



10. References

1. D.Samaržija, N.Antunac, J.L. Havranek. Taxonomy, physiology and growth of *Lactococcus lactis*: a review. *Mljekarstvo* 51 (1) 35-48, 2001.
2. K.H. Schleifer et al. Transfer of *Streptococcus lactis* and Related Streptococci to the Genus *Lactococcus* gen. nov. System. Appl. Microbiol. 6, 183-195 (1985)
3. P. M. F. SHATTOCK AND A. T. R. MATTICK. THE SEROLOGICAL GROUPING OF STREPTOCOCCUS LACTIS (GROUP N) AND ITS RELATIONSHIP TO STREPTOCOCCUS FAECALIS .
4. J.M. Hardie and R.A. Whiley. Classification and overview of the genera *Streptococcus* and *Enterococcus*. *Journal of Applied Microbiology Symposium Supplement* 1997, 83, 1S-11S
5. Е.П. Мирошникова «Микробиология молока и молочных продуктов» 2005.
6. S. L. Cho et al. *Lactococcus chungangensis* sp. nov., a lactic acid bacterium isolated from activated sludge foam. *International Journal of Systematic and Evolutionary Microbiology* (2008), 58, 1844-1849
7. Montville et al. *FOOD MICROBIOLOGY An Introduction*. Second edition. Chapter 18. ASM Press American Society for Microbiology (2008)
8. P.M. Moraes et al. Comparison of phenotypic and molecular tests to identify lactic acid bacteria. *Brazilian Journal of Microbiology* 44, 1, 109-112 (2013)

10. References

9. M. Sakaki et al. Immunological and electrophoretic study of the proteolytic enzymes from various *Lactococcus* and *Lactobacillus* strains. *Journal of Dairy Research* (1995) 62 611-620
10. Vendrel et al. *Lactococcus garvieae* in fish: A review. *Comparative Immunology, Microbiology & Infectious Diseases* 29 (2006) 177-198
11. S. Altun et al. Genotyping of *Lactococcus garvieae* strains from rainbow trout (*Oncorhynchus mykiss*) by 16s rDNA sequencing. *Bull. Eur. Ass. Fish Pathol.*, 24(2) 2004, 119
12. M. Nomura et al. Rapid PCR-Based Method Which Can Determine Both Phenotype and Genotype of *Lactococcus lactis* Subspecies. *APPLIED AND ENVIRONMENTAL MICROBIOLOGY*, Vol. 68, No. 5, p. 2209-2213, May 2002
13. J. L. W. Rademaker et al. Diversity Analysis of Dairy and Nondairy *Lactococcus lactis* Isolates, Using a Novel Multilocus Sequence Analysis Scheme and (GTG)₅-PCR Fingerprinting. *APPLIED AND ENVIRONMENTAL MICROBIOLOGY*, Vol. 73, No. 22, p. 7128-7137, Nov. 2007
14. P. Nieto-Arribas et al. Genotypic and technological characterization of *Lactococcus lactis* isolates involved in processing of artisanal Manchego cheese. *Journal of Applied Microbiology* 107 (2009) 1505-1517

10. References

15. E. Ferná'ndez et al. Comparative Phenotypic and Molecular Genetic Profiling of Wild *Lactococcus lactis* subsp. *lactis* Strains of the *L. lactis* subsp. *lactis* and *L. lactis* subsp. *cremoris* Genotypes, Isolated from Starter-Free Cheeses Made of Raw Milk. *APPLIED AND ENVIRONMENTAL MICROBIOLOGY*, Vol. 77, No. 15, p. 5324-5335, Aug. 2011
16. *I. Boucher et al. Novel Food-Grade Plasmid Vector Based on Melibiose Fermentation for the Genetic Engineering of *Lactococcus lactis*. *APPLIED AND ENVIRONMENTAL MICROBIOLOGY*, Vol 68, No. 12, p. 6152-6161, Dec. 2002
17. L. G. Stoyanova et al. Isolation and Identification of New Nisin-producing *Lactococcus lactis* subsp. *Lactis* from Milk. *Applied Biochemistry and Microbiology*, 2006, Vol. 42, No. 5, pp. 492-499.
18. Bahey-El-Din M, Gahan CG, Griffin BT. *Lactococcus lactis* as a cell factory for delivery of therapeutic proteins. *Current Gene Therapy*. 2010 Feb;10(1):34-45.
19. K. Robinson et al. Mucosal and Cellular Immune Responses Elicited by Recombinant *Lactococcus lactis* Strains Expressing Tetanus Toxin Fragment C. *INFECTION AND IMMUNITY*, Vol 72, No. 5, p. 2753-2761, May 2004
20. J.V. Hernandez et al. Targeting diseases with genetically engineered *Lactococcus lactis* and its course towards medical translation. *Expert Opin. Biol. Ther.* (2011) 11(3):261-267

10. References

21. S. B. Hanniffy et al. Mucosal Delivery of a Pneumococcal Vaccine Using *Lactococcus lactis* Affords Protection against Respiratory Infection. *The Journal of Infectious Diseases* 2007; 195:185-93.
22. G. F. Asensi et al. Oral immunization with *Lactococcus lactis* secreting attenuated recombinant staphylococcal enterotoxin B induces a protective immuneresponse in a murine model. *Microbial Cell Factories* 2013, 12:32
23. M. Medina et al. *Lactococcus lactis* as an adjuvant and delivery vehicle of antigens against pneumococcal respiratory infections. *Bioengineered Bugs* 1:5, 313-325; 2010
24. X.Z. Zhang et al. Expression of *Helicobacter pylori* hspA Gene in *Lactococcus lactis* NICE System and Experimental Study on Its Immunoreactivity. Hindawi Publishing Corporation *Gastroenterology Research and Practice* Volume 2015, Article ID 750932, 6 pages
25. Q. Gu et al. Oral vaccination of mice against *Helicobacter pylori* with recombinant *Lactococcus lactis* expressing urease subunit B. *FEMS Immunol Med Microbiol* 56 (2009) 197-203
26. Bahey-El-Din M, Gahan CG, Griffin BT. *Lactococcus lactis* as a cell factory for delivery of therapeutic proteins. *Current Gene Therapy*. 2010 Feb;10(1):34-45.

10. References

Links:

1. <https://en.wikipedia.org/wiki/Lactococcus>
2. <http://genome.jgi.doe.gov/laccr/laccr.home.html>
3. <http://textbookofbacteriology.net/lactics.html>

